

# Portable Raman Spectroscopy for the Study of Polymorphs and Monitoring Polymorphic Transitions

## INTRODUCTION

Materials can exist in different polymorphic forms, meaning that their crystal structure can vary, even though they have the same chemical composition. Polymorphs exist in organic as well as inorganic materials, including metal oxides and silica (quartz). Different polymorphs have different stabilities and may be formed preferentially based on the crystallization process. Polymorphism is important in pharmaceutical products because the efficacy of a drug can be impacted substantially based on the solubility of the different crystal forms.

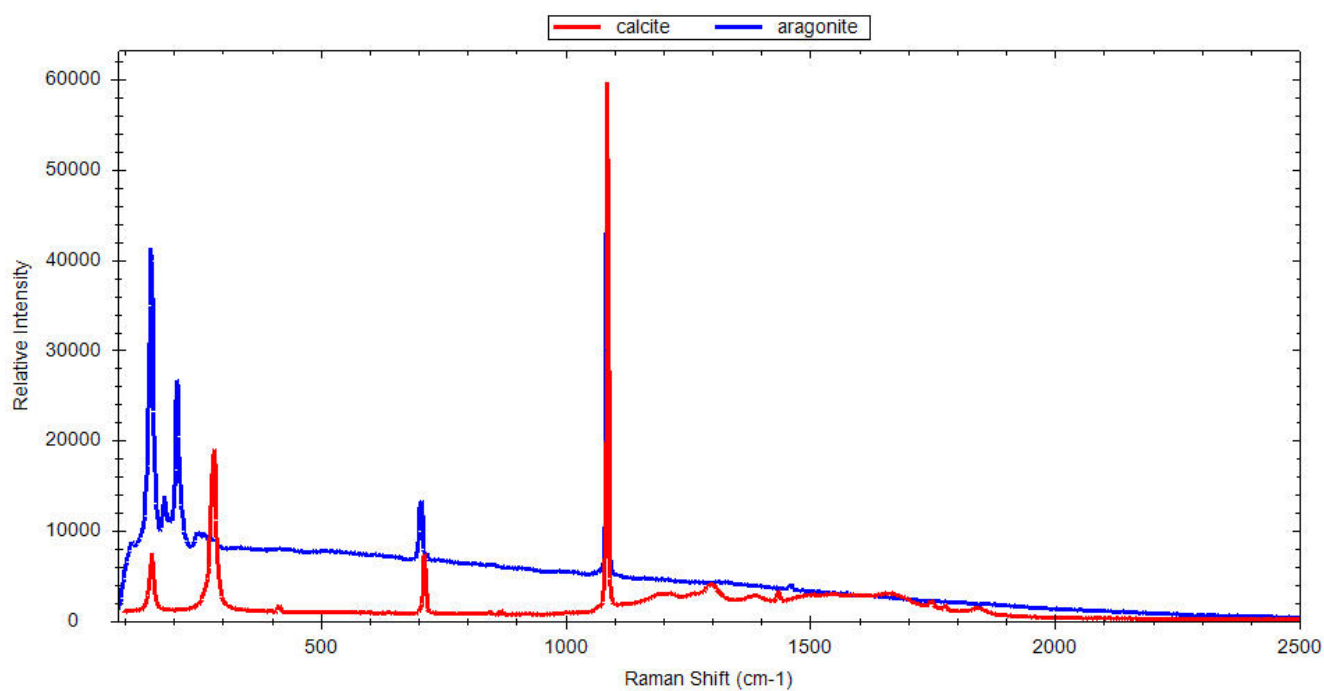
Raman spectroscopy is used for material characterization by analyzing molecular or crystal symmetrical vibrations and rotations that are excited by a laser, and exhibit vibrations specific to the molecular bonds and crystal arrangements in the molecules. Due to its excellent spectral specificity, Raman technology is a valuable tool in distinguishing different polymorphs, and can also be used in the study of

solvate forms, as well as the kinetics of polymorphic transitions and crystallization processes[1-4].

Raman can be used for in situ measurements as a PAT (process analytical technology) tool for online continuous monitoring of chemical reactions as well as polymorph transformations. Portable Raman systems are an excellent tool in the rapid identification of polymorphs that exhibit distinct spectral differences due to the different arrangement of molecules in the crystal. Portable Raman is especially beneficial in process development where polymorphic screening, stability and formation are determined because of its small compact scale and ease of installation and use.

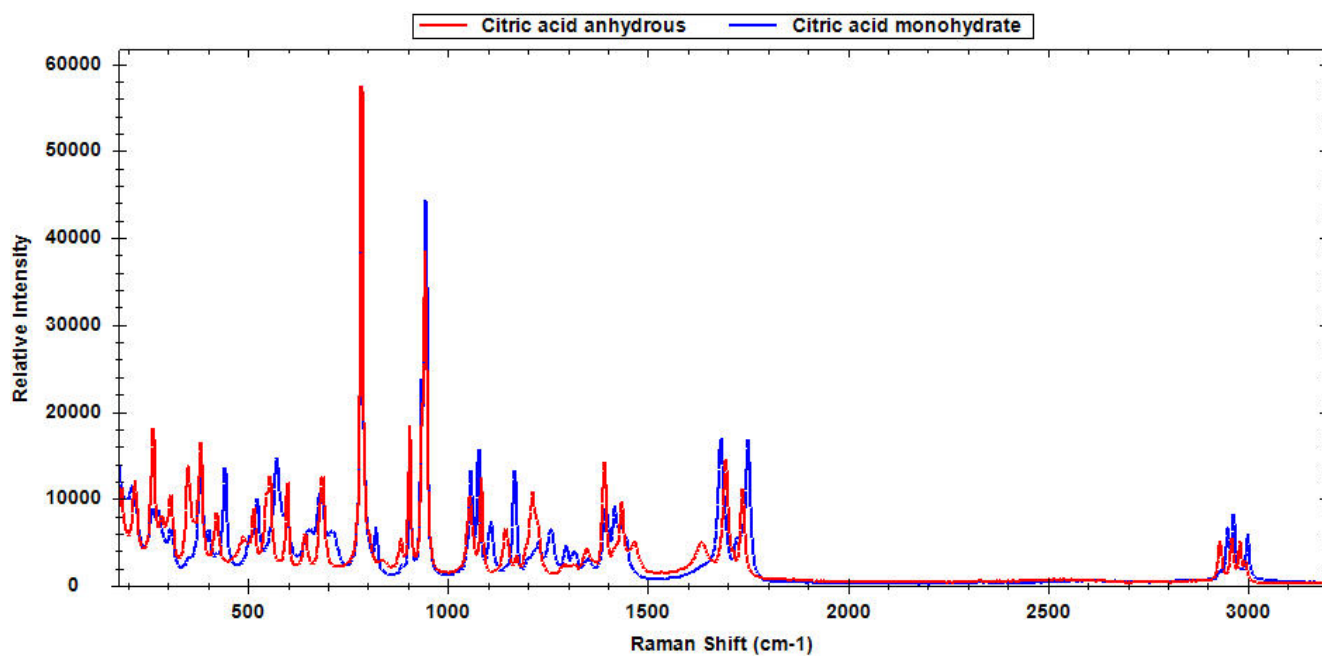
The Raman spectra of several polymorphs are shown in **Figures 1-3** to illustrate how distinct the Raman spectra are for these polymorphic pairs: calcium carbonate (aragonite and calcite), citric acid, and dextrose.

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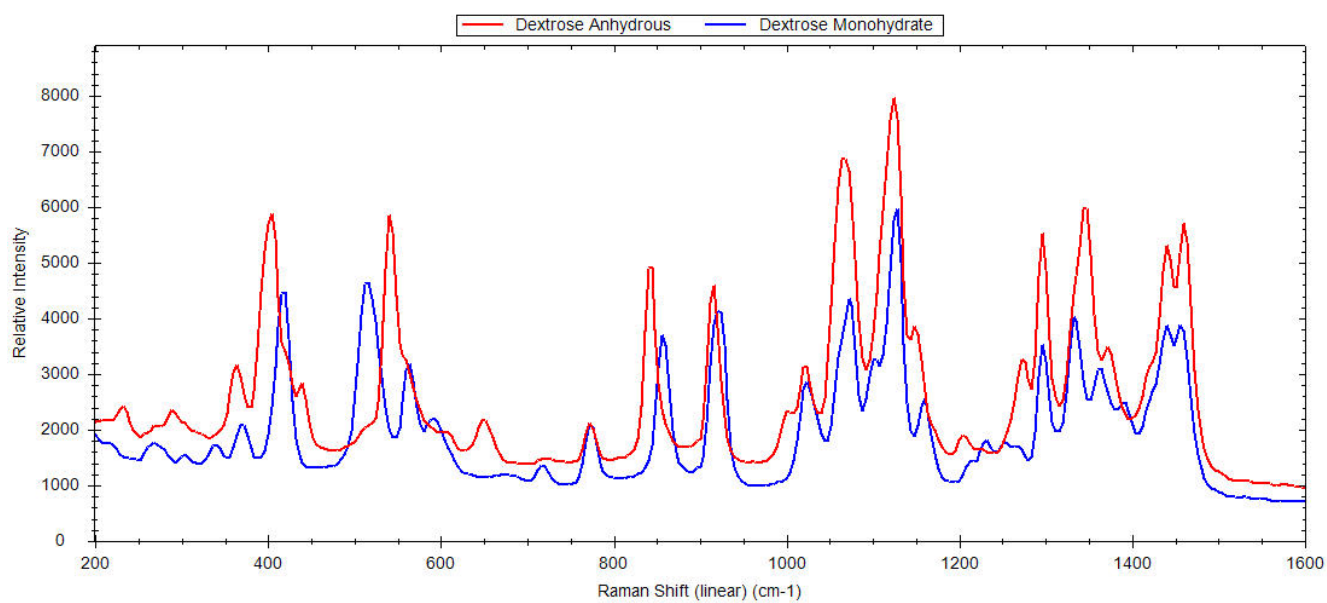


**Figure 1.** Raman spectra of two polymorphs of calcium carbonate: calcite and aragonite

## INTRODUCTION



**Figure 2.** Raman spectra of citric acid polymorphs



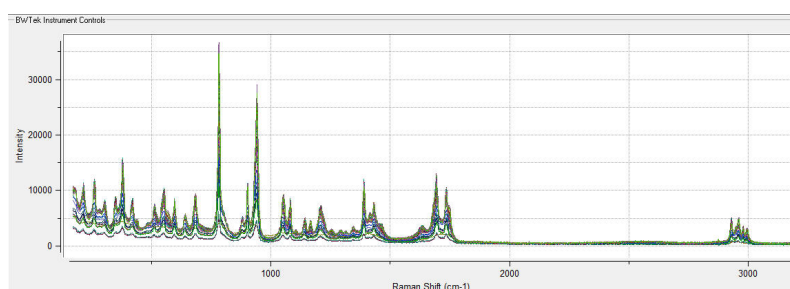
**Figure 3.** Raman spectra of anhydrous dextrose and dextrose monohydrate

## EXPERIMENT

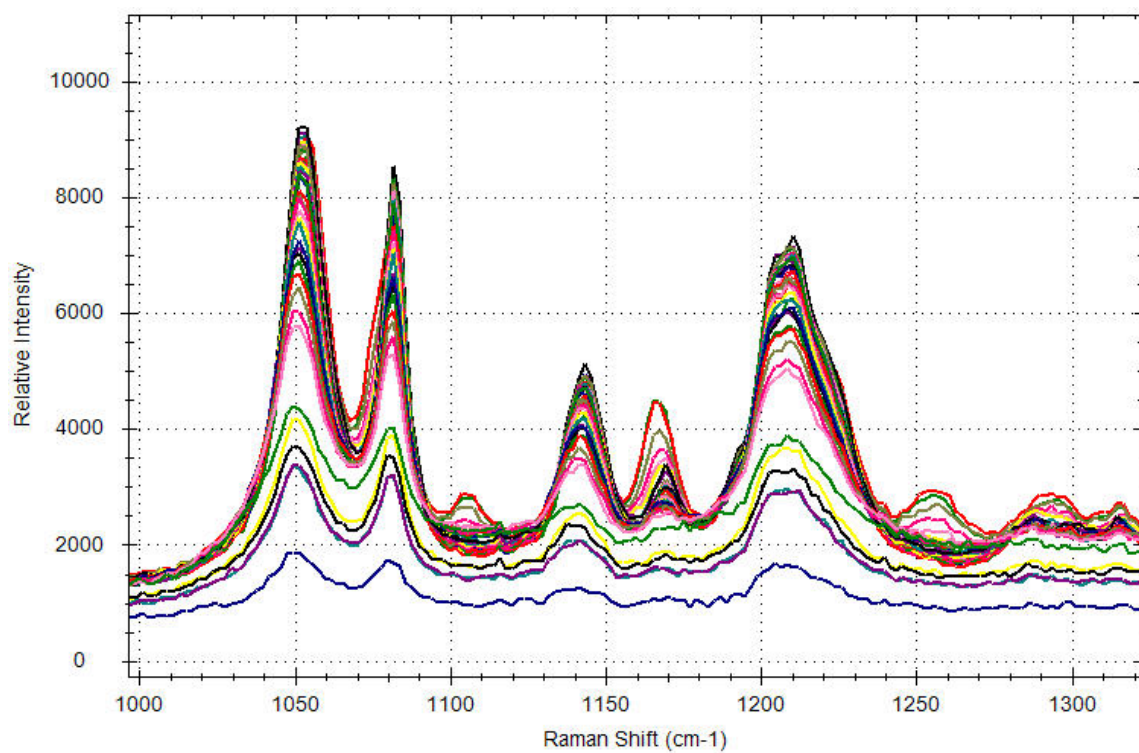
In this work the capability of portable Raman as a process monitoring tool is shown based on the use of B&W Tek's i-Raman Plus. Measurements were performed using portable i-Raman Plus equipped with a sensitive TE-cooled back-thinned CCD as well as a patented CleanLaze® laser excited at 785 nm with 300 mW maximum power output, covering the spectral range from 175-3200  $\text{cm}^{-1}$ . A long shaft lab grade Raman probe is positioned above the sample surface, at a working distance of 5 mm such that the laser is well focused as a spot. Data were collected with an acquisition time of 15-30 seconds with 300 mW laser power.

Citric acid, a well-known food additive, is selected here as a model system to study polymorphism based on two of its solid crystal phases: monohydrate and anhydrous, which crystallize from water at different temperatures[4]. Citric acid monohydrate and citric acid anhydrous were purchased from Sigma-Aldrich. The transition of the monohydrate form to the anhydrous was performed by heating the solid powder from room temperature to 80 ° C.

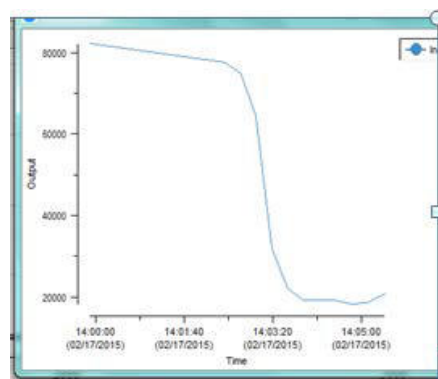
Real-time monitoring and trending of the polymorphic transition of citric acid was done using B&W Tek's BWSP-21pt11 software. This allows for continuous data collection and trending based on evolving PCA scores, chemometric models, or data trends. As the interest here is in following the transition (not quantitating the amount of the forms in the system during this process) we used the trend of the disappearance of monohydrate peak at 1108  $\text{cm}^{-1}$  and formation of the anhydrous form with the change in intensity of the new peak at 1146  $\text{cm}^{-1}$ . There are other peaks specific to the monohydrate form including at 442, 820, 1167, 1260 and 2950  $\text{cm}^{-1}$  and for the anhydrous form at 1635, 2932 and 2982  $\text{cm}^{-1}$  that could be readily used to trend the transition. An overlay of spectra collected continuously as the temperature is increased to 80 ° C is given in **Figure 4**. In the expanded view of the spectral region used for peak trending, the spectral changes on phase transition are evident in **Figure 5**. The trend of the monohydrate peak disappearance that was generated in real-time during data collection is given in **Figure 6**.



**Figure 4.** Overlay of Raman spectra collected every 15 seconds during temperature increase to induce monohydrate to anhydrous transition in citric acid



**Figure 5.** Expanded view of Raman spectra collected during citric acid phase transition

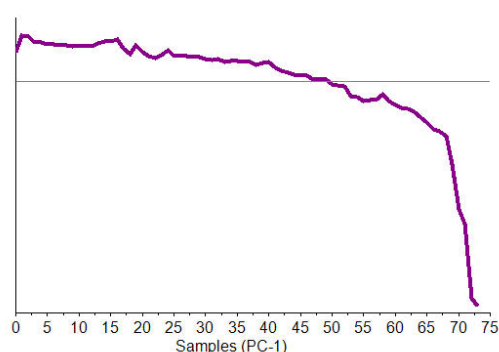


**Figure 6.** Trend of Raman peak area of 1108 cm<sup>-1</sup> citric acid monohydrate, during temperature increase from room temperature to 80 °C

## CONCLUSIONS

Because the spectral changes related to the change from the monohydrate to the anhydrous citric acid are not limited to discrete changes, a more holistic approach using multivariate data analysis, reflecting the systematic spectral changes with the increase in temperature can be captured using Principal component analysis (PCA). Using PCA analysis over the full spectral range, it is found that the first

principal component explains 90% of the data variance over the course of the 75 spectra collected. A line plot of the score of PC-1 vs. sample spectrum shows the same trend as seen by following a single peak, and reflects changes across the Raman spectrum with the conversion for the monohydrate to anhydrous form. **Figure 7** shows the plot of the PC-1 scores.



**Figure 7.** PC-1 score plot from full spectral range PCA analysis of 75 spectra collected during temperature experiment

Here we show some examples of the value of Raman spectroscopy to monitor and identify different polymorphs. We used the portable i-Raman Plus with BWSP-21pt11 software for continuous

monitoring and trending of the monohydrate to anhydrous transition of citric acid, with spectra collected every 15 seconds as the temperature was increased.

## REFERENCES

1. E. Smith and G. Dent, Modern Raman Spectroscopy - A Practical Approach, John Wiley and Sons, Hoboken, NJ, 2005.
2. J. Huang and M. Dali, J. Pharm. and Biomedical Anal. 86(2013) 92-99.
3. M. Steindl et al, Chem. Eng. and Processing 44(2005) 471-475.
4. A. Caillet, F. Puell, G. Fevotte, Chem. Eng. and Processing 47(2008) 377-382.

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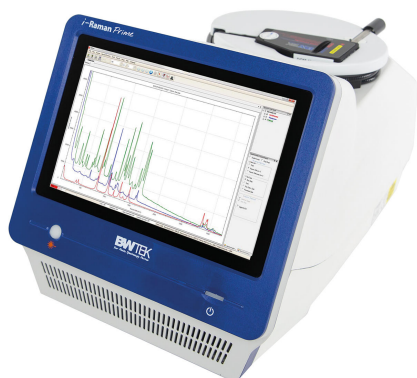
## CONFIGURATION



### i-Raman Plus 785S

i-Raman® Plus 785S は、弊社の革新的かつインテリシエントなスペクトロメータテクノロジーを備えた i-Raman ホータフル型ラマンスペクトロメータの受賞歴のあるシリーズの構成要素です。このホータフル型ラマンスペクトロメータは、高い量子効率、TE 冷却、ならびに高いダイナミックレンジを備えた CCD アレイ検出器を用い、積分時間でさえ最長30分、低ノイズの傑出した性能を提供します。こうして、弱いラマン信号も測定することかできます。

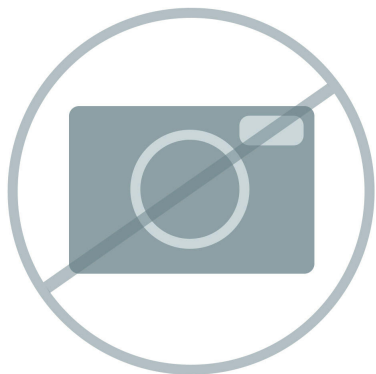
i-Raman Plus 785S は、 $65\text{ cm}^{-1}$  から  $3350\text{ cm}^{-1}$  までの測定を可能にするコンフィクレーションを有する幅広いスペクトル範囲と高分解能のユニークな組み合わせを特徴としています。システムの小さな底面、軽量構造、そして低消費電力により、とこてもラマン分析を研究レベルで実施することかできます。i-Raman Plus には、サンプル採取を簡単にする光ファイバーフローが装備されており、キューベットホルター、ヒテオマイクロスコフ、フローホルター付き XYZ スライドテーブル、ならびに弊社独自の多変量解析ソフトウェア BWIQ® および同定ソフトウェア BWID® と共に使用することかできます。i-Raman Plus により、品質分析および定量分析のための高精度のラマンソリューションを常に使用することかできます。



### i-Raman Prime 785S

i-Raman® Prime 785S は、タブレット PC および光ファイバーサンプリング内蔵の、低ノイズかつ高スループットの完全一体型のラマンシステムです。このポータブル型ラマン分光計は、高い量子効率、TE 冷却 (-25 ° C)、ならびに高いダイナミックレンジを備えた CCD アレイ検出器を使用し、リアルタイムでの定量化と同定を含む研究レベルでのラマン分析を提供します。高スループットにより、傑出した信号対雑音比のラマンスペクトルを得ることのできるため、速いプロセスの測定、および最も弱いラマン信号でさえも可能となり、サンプルの微細な相違も検出できます。

i-Raman Prime 785S には、そのポータブル式の構造に加え、幅広いスペクトル領域と高い分解能というユニークな組み合わせが装備されているため、 $150\text{ cm}^{-1}$  から  $3350\text{ cm}^{-1}$  までの測定が可能です。i-Raman Prime はバッテリーでの稼働が可能なので、容易に持ち運ぶことができます。そのため、場所を選ばず、高精度かつ質的、量的に価値の高いラマン分析を研究レベルで実施することできます。システムは、不透明なハッチャーシ越しの分析用の、弊社 STRaman® テクノロジーの使用向けに最適化されました。



532 nm または 785 nm の励起波長の ラマンフロー BAC100/BAC102 のためのオプションとしての浸漬シャフト。ハーフオーロム製 O リングで密封された石英ガラスウィンドウ付き、外径 12.0 mm のステンレス 316L 製 76.2 mm 長のホチキスを有しています。空中での作動距離 5.0 mm。-55 ° C から 200 ° C で使用可能。ハッキングは、低圧 (< 1 bar) では液密です。