



Application Note AN-RS-048

# Phosphates speciation with Raman spectroscopy

## A simpler alternative to wet chemical analysis methods

Phosphates are integral to a wide range of industrial processes, from agriculture and food production to water treatment and pharmaceuticals. The ability to accurately identify and monitor the transformation of different phosphate species—such as phosphoric acid ( $\text{H}_3\text{PO}_4$ ), dihydrogen phosphate ( $\text{H}_2\text{PO}_4^-$ ), hydrogen phosphate ( $\text{HPO}_4^{2-}$ ), and phosphate ( $\text{PO}_4^{3-}$ )—is essential for optimizing these processes and ensuring

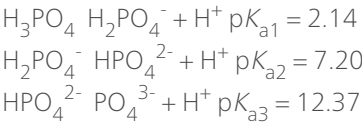
compliance with regulatory standards. Each phosphate ion species exhibits unique chemical properties and behaviors, influencing their reactivity, solubility, response to excitation, and roles in industrial applications.

This Application Note demonstrates the ability of Metrohm's MIRA XTR handheld Raman instrument to speciate phosphate ions.

INTRODUCTION

Raman spectroscopy enables non-contact, real-time monitoring of complex chemical systems. Raman offers users exceptional sampling ease and accuracy, especially in harsh environments like strong acid testing. Its high specificity and sensitivity to small structural changes in a molecule make it an ideal sensitive tool for speciation. This application describes the use of handheld Raman to monitor phosphate species throughout an acid/base titration [1]. The transformation of phosphate species from phosphoric acid through its deprotonated forms to the simple phosphate ion can be monitored by

Raman spectroscopy. The protonation state of H<sub>3</sub>PO<sub>4</sub> significantly impacts industrial manufacturing processes and applications such as fertilizer application, wastewater treatment, and corrosion control [2].



Understanding and tracking these changes optimizes the use of phosphates, prevents unwanted side reactions, and maintains process stability.

METHOD

Raman spectroscopy was used to monitor phosphate species directly throughout the acid/base titration of phosphoric acid.  
**Sample preparation:** A experimental solution of 2% phosphoric acid (v/v) was prepared from a 10%

phosphoric acid stock solution (Sigma-Aldrich) with deionized water. The titrant was a 5 mol/L NaOH solution prepared by diluting 10 mol/L NaOH with deionized water.

Table 1. Titration and handheld Raman equipment and reagents.

| Equipment                                                                                   |
|---------------------------------------------------------------------------------------------|
| <b>Titration</b><br>907 Titrando<br>tiamo 2.5 software<br>801 stirrer<br>800 dosino (20 mL) |
| <b>Raman</b><br>MIRA XTR with XLWD attachment<br>MIRA Cal DS software                       |
| <b>Reagents</b><br>2% phosphoric acid<br>5 mol/L NaOH<br>Deionized water                    |

## MEASUREMENT

A 907 Titrand automatic titrator (Figure 1) was used to titrate the 2% phosphoric acid against standardized 5 mol/L NaOH solutions. At each set point, the pH of the sample was measured, and the Raman spectrum was collected with MIRA XTR (Figure 1).

Raman data was collected from the top surface of the solution. Alternatively, Raman data can also be acquired directly from the solution using an immersion attachment or through the titration vessel's glass wall.



**Figure 1.** The 907 Titrand (top) and MIRA XTR (bottom) from Metrohm were used for simultaneous collection of pH and Raman data.

**Table 2.** MIRA XTR operational presets used for this study.

| Parameter        | Setting |
|------------------|---------|
| Laser power      | 50 mW   |
| Integration time | 30 s    |
| Averages         | 3       |
| Raster           | OFF     |

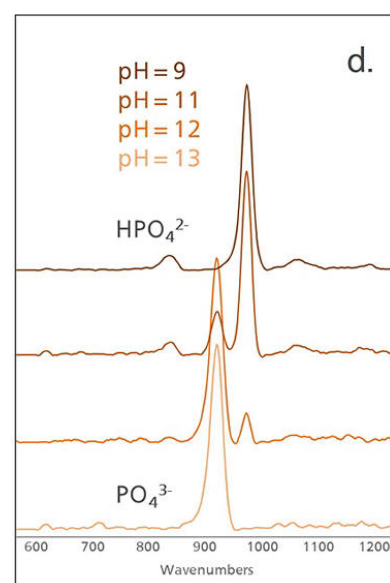
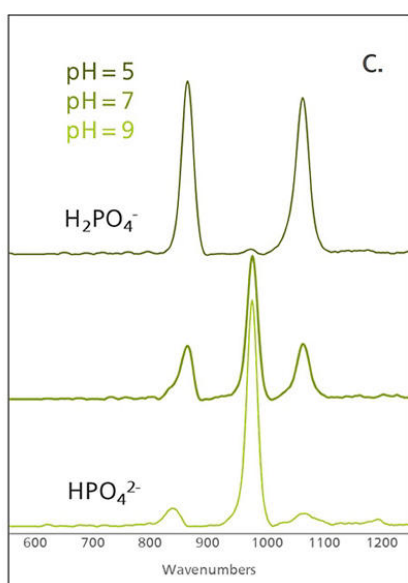
## RESULTS

The first, second, and third midpoints of phosphoric acid occur at approximately pH 2, pH 7, and pH 12, respectively (Figure 2a). These correspond closely to the  $pK_a$  values for each deprotonation step of phosphoric acid. Likewise, the first and second equivalence points at approximately pH 4 and pH 9 are followed by complete deprotonation (Figure 2a). Raman spectra at the different equivalence points displayed distinct differences (Figure 2b–d).

Figure 2b shows the changes that occur between pH 1–5. The peak at  $890\text{ cm}^{-1}$ , associated with the symmetric stretch  $\nu_s(\text{P}(\text{OH})_3)$  of  $\text{H}_3\text{PO}_4$ , shifts to  $876\text{ cm}^{-1}$ , which corresponds to the symmetric stretch  $\nu_s(\text{P}(\text{OH})_2)$  of  $\text{H}_2\text{PO}_4^-$ . The peak at  $1078\text{ cm}^{-1}$ ,

attributed to the symmetric stretching  $\nu_s(\text{PO}_2)$ , gradually increases [1].

As the titration progresses from pH 5 to 9 (Figure 2c), the peaks at  $876$  and  $1078\text{ cm}^{-1}$  (associated with  $\text{H}_2\text{PO}_4^-$ ) gradually decrease, and a new peak appears at  $990\text{ cm}^{-1}$ , attributed to the symmetric stretch  $\nu_s(\text{PO}_3)$  of  $\text{HPO}_4^{2-}$ . Finally, as the pH increases from 9 to 13 (Figure 2d), the  $990\text{ cm}^{-1}$  peak gradually disappears, and a new peak at  $937\text{ cm}^{-1}$  emerges, attributed to the symmetric stretching mode  $\nu_s(\text{PO}_4)$ . These results demonstrate that Raman spectroscopy can effectively track changes in phosphate protonation states during titration reactions.



**Figure 2.** a) Titration curve of the 2% phosphoric acid against 5 mol/L NaOH solution. Raman spectra of the titration at the full range of b) pH 1–5, c) pH 5–9, and d) pH 9–13 showing the spectra of different species at different deprotonation points.

## CONCLUSION

Raman spectroscopy provides real-time analysis of the analyte species present in solution, permitting precise identification of phosphate ions as they transition with pH changes. A small handheld Raman system like MIRA XTR achieves fast and direct confirmation of wet chemical techniques like titration with no need for

reagents or complex sample preparation. Additionally, it enables continuous monitoring of dynamic systems, providing detailed and accurate insight into the speciation process and reducing the potential for error in measurements.

## REFERENCES

1. Lackey, H. E.; Nelson, G. L.; Lines, A. M.; et al. Reimagining PH Measurement: Utilizing Raman Spectroscopy for Enhanced Accuracy in Phosphoric Acid Systems. *Anal. Chem.* **2020**, *92* (8), 5882–5889.  
[DOI:10.1021/acs.analchem.9b05708](https://doi.org/10.1021/acs.analchem.9b05708)
2. *Determination of phosphoric acid with sodium hydroxide.*  
<https://www.metrohm.com/en/applications/application-notes/aa-t-001-100/an-t-237.html>  
(accessed 2025-02-03).

## CONTACT

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## CONFIGURATION



### MIRA XTR Advanced

MIRA XTR est une alternative pour les systèmes haute puissance 1 064 nm. Piloté par un traitement informatique avancé, MIRA XTR utilise un laser de 785 nm plus sensible ainsi que des algorithmes XTR pour extraire les données Raman de la fluorescence de l'échantillon. MIRA XTR dispose également d'un balayage de trame orbital (ORS, Orbital Raster Scanning) pour fournir une meilleure couverture de l'échantillon, améliorant ainsi l'exactitude des résultats.

Le pack MIRA XTR Advanced comprend un standard de calibrage, un embout universel intelligent, un embout à angle droit, un support de flacon et des accessoires SERS MIRA. Un package complet pour tous les types d'analyse. Fonctionnement en classe 3B. MIRA XTR prend en charge les bibliothèques Raman de Metrohm portables.