

850 Professional IC



Anion MSM-HC HP-Gradient–2.850.2260

Manual

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1 Introduction

1.1 Instrument description

The instrument **850 Professional IC Anion MSM-HC HP-Gradient** (2.850.2260) is one of the model versions of the Professional IC line of instruments manufactured by the Metrohm Company. The Professional IC line of instruments is distinguished by:

- the **intelligence** of its components, which are able to monitor and optimize all functions and to provide documentation according to FDA requirements.
- its **compactness**.
- its **flexibility**. A suitable model version exists for every application. Individual instruments can be converted, expanded or modified to create a different instrument as needed.
- its **transparency**. All components are easily accessible and arranged in a clear manner.
- its **safety**. Chemicals and electronics are separated and a leak sensor is integrated in the wet end.
- its **environmental compatibility**.
- **low noise emission**.

The instrument is operated with **MagIC Net** software. It is connected via a USB connection to a PC on which MagIC Net is installed. The software automatically recognizes the instrument and checks its functional readiness. MagIC Net controls and monitors the instrument, evaluates the measured data and administers it in a database. The operation of MagIC Net is described in the online help or in the tutorial for MagIC Net.

The instrument contains the following components:

Eluent degasser

The eluent degasser removes gas bubbles and dissolved gases from the eluent. For degassing, the eluent flows into a vacuum chamber through a special fluoropolymer capillary.

High pressure pump

The intelligent and low pulsation high pressure pump pumps the eluent through the system. It is equipped with a chip on which its technical specifications and "life history" (operating hours, service data, ...) are saved.

Inline filter

Inline filters protect the separation column securely against possible contamination from the eluent. Inline filters can however also just as well be



used for the purpose of protecting other sensitive components against contaminations in the solutions used. The filter platelets with a pore size of 2 μm can be replaced quickly and easily. They remove particles like e. g. bacteria and algae from the solutions.

Pulsation absorber

The pulsation absorber protects the separation column from damage caused by pressure fluctuations when switching the injection valve, and reduces interfering pulsations during highly sensitive measurements.

Sample degasser

The sample degasser removes gas bubbles and dissolved gases from the sample. For degassing, the sample flows into a vacuum chamber through a special fluoropolymer capillary.

Injection valve

The injection valve connects the eluent and sample path through rapid and precise valve switchover. A precisely measured amount of sample solution is injected and rinsed with eluent onto the separation column.

Column thermostat

The column thermostat controls the temperature of the column and eluent channel and thus ensures stable measuring conditions. It provides space for 2 separation columns.

High Capacity Metrohm Suppressor Module (MSM-HC)

The MSM-HC is used for chemical suppression with concentrated eluents during anion analysis. It is pressure-stable, robust and resistant to solvents.

Peristaltic pump

The Peristaltic pump is used for pumping sample and auxiliary solutions. It can rotate in both directions.

Separation column

The intelligent separation column is the heart of the ion chromatographic analysis. It separates the different components corresponding to their interactions with the column. Metrohm separation columns are equipped with a chip on which their technical specifications and their history (first use / setting up, operating hours, injections, ...) are saved.

1.2 Intended use

If required, the instrument can also be used for the determination of cations and anions without suppression.

This instrument is suitable for processing chemicals and flammable samples. The usage of the 850 Professional IC – Anion MSM-HC HP-Gradient therefore requires that the user has basic knowledge and experience in the handling of toxic and caustic substances. Knowledge with respect to the application of the fire prevention measures prescribed for laboratories is also mandatory.

1.3 About the documentation



Please read through this documentation carefully before putting the instrument into operation. The documentation contains information and warnings which the user must follow in order to ensure safe operation of the instrument.

1.3.1 Symbols and conventions

The following symbols and styles are used in this documentation:

(5-12)	Cross-reference to figure legend The first number refers to the figure number, the second to the instrument part in the figure.
1	Instruction step Carry out these steps in the sequence shown.
	Warning This symbol draws attention to a possible life hazard or risk of injury.
	Warning This symbol draws attention to a possible hazard due to electrical current.

A yellow triangular warning sign with a black border, featuring three wavy vertical lines and a horizontal line at the bottom, indicating a hot surface.

WARNING

WARNING

Only personnel qualified by Metrohm are authorized to carry out service work on electronic components.



WARNING

Never open the housing of the instrument. The instrument could be damaged by this. There is also a risk of serious injury if live components are touched.

There are no parts inside the housing which can be serviced or replaced by the user.

Supply voltage



WARNING

An incorrect supply voltage can damage the instrument.

Only operate this instrument with a supply voltage specified for it (see rear panel of the instrument).

Protection against electrostatic charges



WARNING

Electronic components are sensitive to electrostatic charges and can be destroyed by discharges.

Do not fail to pull the power cord out of the power socket before you set up or disconnect electrical plug connections at the rear of the instrument.

1.4.3 Tubing and capillary connections



CAUTION

Leaks in tubing and capillary connections are a safety risk. Tighten all connections well by hand. Avoid applying excessive force to tubing connections. Damaged tubing ends lead to leakage. Appropriate tools can be used to loosen connections.

Check the connections regularly for leakage. If the instrument is used mainly in unattended operation, then weekly inspections are mandatory.



1.4.4 Flammable solvents and chemicals

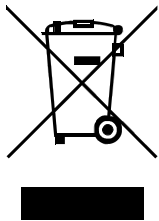


WARNING

All relevant safety measures are to be observed when working with flammable solvents and chemicals.

- Set up the instrument in a well-ventilated location (e.g. fume cupboard).
- Keep all sources of flame far from the workplace.
- Clean up spilled liquids and solids immediately.
- Follow the safety instructions of the chemical manufacturer.

1.4.5 Recycling and disposal



This product is covered by European Directive 2012/19/EU, WEEE – Waste Electrical and Electronic Equipment.

The correct disposal of your old instrument will help to prevent negative effects on the environment and public health.

More details about the disposal of your old instrument can be obtained from your local authorities, from waste disposal companies or from your local dealer.

2 Overview of the instrument

2.1 Front

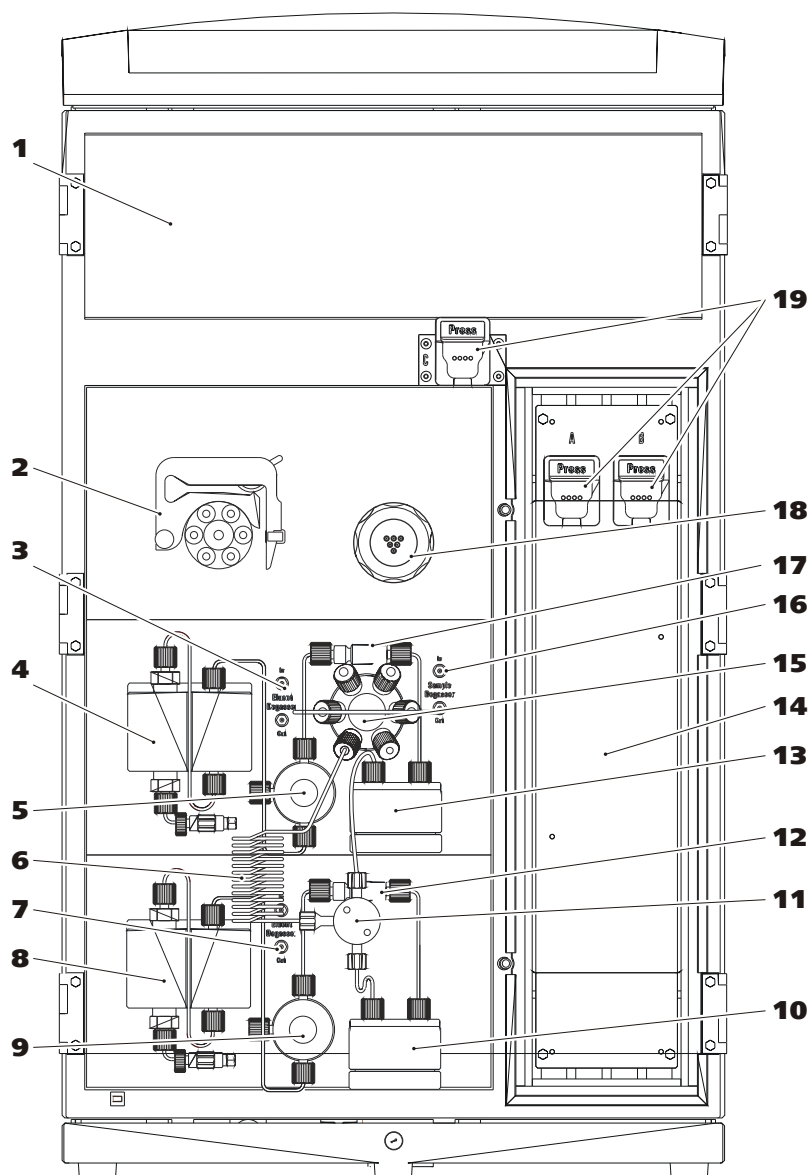


Figure 1 Front 850 Professional IC Anion MSM-HC HP-Gradient

1 Detector chamber

Space for the detector. (The detector must be ordered separately).

2 Peristaltic pump

See Chapter 3.17, Page 50.

3 Eluent degasser

See Chapter 3.9, Page 32.

4 High pressure pump

See Chapter 3.10, Page 34.

- | | | | |
|-----------|---|-----------|---|
| 5 | Purge valve
For deaerating the high pressure pump <i>See Chapter 3.10.1, Page 34.</i> | 6 | Mixing coil for high pressure gradients (6.2758.000) |
| 7 | Eluent degasser
For two additional eluents (<i>see chapter 3.9, page 32</i>). | 8 | High pressure pump
<i>See Chapter 3.10, Page 34.</i> |
| 9 | Purge valve
For deaerating the high pressure pump (<i>see chapter 3.10.1, page 34</i>). | 10 | Pulsation damper
<i>See Chapter 3.12, Page 39.</i> |
| 11 | T connection (6.2744.150) | 12 | Inline filter
<i>See Chapter 3.11, Page 38.</i> |
| 13 | Pulsation damper
<i>See Chapter 3.12, Page 39.</i> | 14 | Column thermostat
<i>See Chapter 3.15, Page 44.</i> |
| 15 | Injection valve
<i>See Chapter 3.14, Page 42.</i> | 16 | Sample degasser
<i>See Chapter 3.13, Page 40.</i> Use optional. |
| 17 | Inline filter
<i>See Chapter 3.11, Page 38.</i> | 18 | MSM-HC
<i>See Chapter 3.16, Page 47.</i> |
| 19 | Column holder
For two separation columns (<i>see chapter 3.21, page 58</i>) in the column thermostat and one outside the column thermostat. | | |

2.2 Rear

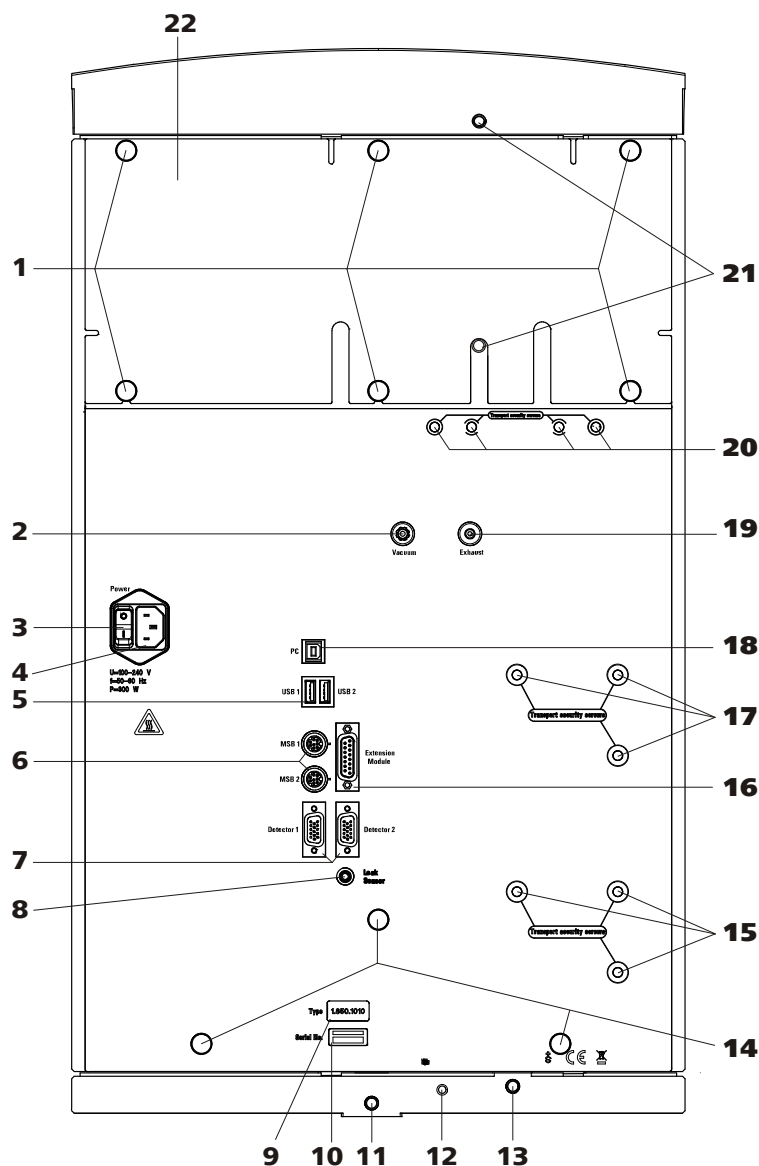


Figure 2 Rear 850 Professional IC Anion MSM-HC HP-Gradient

1 Knurled screws

For fastening the rear panel (2-22) and handle (5-2).

3 Mains switch

For switching the instrument on and off.

I = ON

0 = OFF

2 Vacuum connection

For connecting further degassing chambers in extension modules.

Labeled with *Vacuum*.

4 Mains connection socket

For connecting the mains cable.

- | | | | |
|-----------|--|-----------|--|
| 5 | USB connectors
2 USB connectors (labeled with <i>USB 1</i> and <i>USB 2</i>). | 6 | MSB connectors
2 MSB connectors (labeled with <i>MSB 1</i> and <i>MSB 2</i>) for connecting MSB devices
Caution: The 850 must be switched off when connecting a device.
MSB = Metrohm Serial Bus. |
| 7 | Detector connectors
2 detector connectors (labeled with <i>Detector 1</i> and <i>Detector 2</i> for connecting Metrohm detectors. | 8 | Leak sensor connection socket
For connecting the leak sensor connector plug (7- 2). |
| 9 | Instrument type | 10 | Serial number |
| 11 | Drainage tubing connection
For connecting a drainage tubing 6.1816.020 (8- 8). | 12 | Leak sensor connection cable
For connecting the leak sensor. |
| 13 | Drainage tubing connection
For connecting a drainage tubing 6.1816.020 (8- 9). | 14 | Knurled screws
For fastening the rollers. |
| 15 | Transport locking screws
For securing the lower high pressure pump (17- 4) when transporting the instrument (only necessary for instruments with two high pressure pumps). | 16 | Extension module connection
For connecting an extension module (labeled with <i>Extension Module</i>). |
| 17 | Transport locking screws
For securing the high pressure pump (17- 4) when transporting the instrument. | 18 | PC connection socket
For connecting the instrument to the computer with the USB cable 6.2151.020. |
| 19 | Waste air opening
For extracting the air from the vacuum chamber.
Labeled with <i>Exhaust</i> . | 20 | Transport locking screws
For securing the vacuum pump when transporting the instrument. |
| 21 | Drainage tubing connection
For connecting a drainage tubing 6.1816.020 (8- 1). | 22 | Rear panel
Removable. Access to the detector chamber. |



CAUTION

When connecting an instrument to the MSB connector (2-**6**) you **must** switch off the 850 Professional IC.

3 Installation

3.1 About this chapter

The *Installation* chapter contains

- this overview
- a brief set of instructions for the initial installation of the 850 Professional IC Anion MSM-HC HP-Gradient (see chapter 3.2, page 11). At each step you will find cross-references to more detailed installation instructions for individual components, should you require such aids.
- an installation diagram (see chapter 3.3, page 14), showing a completely installed instrument.
- several chapters with detailed installation instructions for all components, including those that are already installed at the time the instrument is delivered.

3.2 Initial installation



NOTICE

A large number of the capillary connections are already connected at the time the instrument is delivered.

Installing 850 Professional IC Anion MSM-HC HP-Gradient

1 Setting up the instrument

See Chapter 3.4, Page 16.

2 Installations on the rear of the instrument

- Remove handle and rollers (see chapter 3.6.1, page 19).
- Place the detector in the instrument and connect it (see manual of the detector).
- Remove transport locking screws (see "Removing transport locking screws", page 22).
- Connect the leak sensor (see "Connecting the leak sensor", page 22).
- Connect drainage tubings (see "Installing drainage tubings", page 25).



3 Connecting the eluent path

- Equip an eluent aspiration tubing for each of the two high pressure pumps and connect with the eluent bottles (*see chapter 3.8.1, page 28*).
- Instead of the column, connect the 6.2744.040 coupling with a 6.2744.070 short pressure screw at the end of the pre-installed column inlet capillary.
- Use a 6.2744.070 short pressure screw to connect the capillary of the MSM-HC labeled with *in* to the other end of the 6.2744.040 coupling.
- Connect the detector inlet capillary and the capillary of the MSM-HC labeled with *out* to one another using one 6.2744.040 coupling and two 6.2744.070 short pressure screws (*see manual of the detector*).

4 Connecting the sample path



NOTICE

The sample degasser does not have to be connected. We recommend the usage of the sample degasser only if the sample matrix requires it.

- Guide the 6.1803.040 sample aspiration capillary connected to the sample input of the injection valve out of the instrument through a capillary feed-through and connect it with the Sample Processor or
 - *If the sample degasser is used* connect the sample aspiration capillary with a 6.2744.090 long pressure screw to the input of the sample degasser (*see chapter 3.13, page 40*).
- Connect a part of the 6.1803.040 sample aspirating capillary with a 6.2744.090 long pressure screw to the input of the sample degasser. Guide the other end out of the instrument through a capillary feed-through and connect it to the Sample Processor.

5 Installing the peristaltic pump

(see chapter 3.17.2, page 51).

- Connect the 6.1803.020 aspirating capillary for regeneration solutions with a 6.2744.034 tubing olive and a 6.2744.070 short pressure screw to the aspiration end of the 6.1826.320 pump tubing for the regeneration solution.
- Place the pump tubing in a tubing cartridge.

- Connect the 6.1803.020 aspirating capillary for rinsing solutions with a 6.2744.034 tubing olive and a 6.2744.070 short pressure screw to the aspiration end of the second 6.1826.320 pump tubing for the rinsing solution.
- Place the second pump tubing in the second tubing cartridge.
- Place both tubing cartridges into the peristaltic pump.

6 Connecting the MSM-HC

(see chapter 3.16.2, page 47)

- Connect the capillary labeled with *regenerant* with a 6.2744.180 pump tubing connection and a 6.2744.070 short pressure screw to the peristaltic pump on the outlet end of the pump tubing for the regeneration solution.
- Connect the capillary labeled with *Rinsing solution* with a 6.2744.180 pump tubing connection and a 6.2744.070 short pressure screw to the peristaltic pump on the outlet end of the pump tubing for the rinsing solution.
- Guide the two capillaries labeled with *Waste reg.* and *Waste rins.* into a waste container and fasten them there.

7 Connecting the instrument

- Connect the instrument to the PC with a 6.2151.020 USB cable .
- Connect instrument to mains supply .

8 Initial start-up

(see chapter 4.1, page 60)

- Switch on the PC and start MagIC Net.
- Switch on the instrument.
- Deaerate the high pressure pump (see chapter 3.10.2, page 36).
- Set contact pressure of the peristaltic pump (see "Set flow rate", page 54).
- Rinse instrument without column.

9 Installing guard and separation column

- Remove the 6.2744.040 coupling between the column inlet capillary and the MSM-HC capillary labeled with *in*.
- Connect guard column (optional) (see chapter 3.20, page 56)
 - Fasten the guard column to the end of the column inlet capillary according to the specifications contained in the leaflet accompanying the guard column.
 - Rinse guard column.



- Connect the separation column (*see chapter 3.21, page 58*)
 - Fasten the inlet of the separation column either to the end of the column inlet capillary or to the guard column (if used) according to the specifications contained in the leaflet accompanying the column.
 - Fasten the MSM-HC capillary labeled with *in* with a 6.2744.070 PEEK pressure screw at the output of the separation column.

10 Conditioning the instrument

(see "Conditioning the system", page 61)

The instrument is now ready for measuring samples.

3.3 Installation diagram

Figure 3 Installation diagram shows the capillary connections with the application of the gradient technique and chemical suppression (MSM-HC).

The arrangement of the modules in the diagram corresponds to the front view of the instrument. Liquid containers (eluent bottle, sample vessel, waste container, auxiliary solutions container) and guard column (*see chapter 3.20, page 56*) are not shown in the diagram..

Most of the capillaries are already pre-installed at the time the instrument is delivered. Capillaries on which nothing needs to be done at the time of initial installation are not numbered in the diagram.

The diagram illustrates the three main components of a cordless drill, numbered 1 through 4:

- 1. Drill bit:** The pointed tool used for drilling into materials.
- 2. Drill shaft:** The central rod that connects the drill bit to the chuck and handle.
- 3. Drill chuck:** The component that holds the drill bit in place and allows it to rotate.
- 4. Drill handle:** The grip used to operate the drill, shown separately above the main assembly.

Figure 4 Connection of capillaries with pressure screws

- | | |
|--|---|
| <p>1 PEEK pressure screw (6.2744.014)
Use on the injection valve.</p> | <p>2 Connection capillary</p> |
| <p>3 PEEK pressure screw, short (6.2744.070)
For use on the high pressure pump, the purge valve, the inline filter, the pulsation absorber, the guard column and the separation column.</p> | <p>4 PEEK pressure screw, long (6.2744.090)
Use on special components. Is not used on all instruments.</p> |



In order to keep the dead volume as low as possible, capillary connections should generally be as short as possible.



For an improved overview, capillary and tubing connections can be bundled with the 6.1815.010 spiral band.

Connection capillaries

PEEK capillaries and PTFE capillaries are used in the IC system.

PEEK capillaries (poly-
etheretherketone)

PEEK capillaries are temperature-resistant up to 100°C, stable under pressure up to 400 bar, flexible, chemically inert and exhibit an extremely smooth surface. They can be readily cut down to the desired length with the 6.2621.080 capillary cutter.

Usage:

- PEEK capillaries (6.1831.010) with an internal diameter of 0.25 mm for the entire high-pressure section.



- PEEK capillaries (6.1831.030) with an internal diameter of 0.75 mm for sample handling in the ultratrace range.



CAUTION

For the capillary connections between the injection valve and detector , PEEK capillaries with an internal diameter of 0.25 mm must be used. These are already connected to a newly delivered instrument.

PTFE capillaries (polytetrafluoroethylene)

PTFE capillaries are transparent and enable visual tracing of the liquids to be pumped. They are chemically inert, flexible and temperature-resistant up to 80°C.

Usage:

PTFE capillaries (6.1803.0x0) are used for the low-pressure section.

- PTFE capillaries with internal diameter of 0.5 mm for sample handling.
- PTFE capillaries with internal diameter of 0.97 mm for sample handling as well as for rinsing solutions (they do not have to be in the scope of delivery of the instrument).

Capillary connections

In order to achieve optimum analysis results, capillary connections in an IC system must be absolutely tight and free of dead volume. Dead volume occurs if two capillary ends connected to each other do not fit exactly, thus allowing liquid to escape. There are two possible reasons for this:

- The capillaries do not have exactly cut edges.
- The two capillary ends do not completely meet.

One prerequisite for dead volume free capillary connection is, that both capillary ends are cut exactly plane. Therefore we recommend only to cut PEEK capillaries with the capillary cutter (6.2621.080).

Creating dead volume free capillary connections

To create dead volume free capillary connections, proceed as follows:

- 1** Slide the pressure screw over the capillary. Ensure that the capillary protrudes 1–2 mm from the tip of the pressure screw.
- 2** Plug the capillary all the way into the connection or coupling until the stop.
- 3** Only then start turning the pressure screw, while keeping the capillary pressed in space.

The enclosed set of varicolored sleeves for PEEK capillaries (6.2251.000) serves to easily differentiate the various flows of liquid in the system through color coding. Each capillary leading a given liquid (e. g. eluent) can be highlighted with sleeves of the same color.

- 1 Slide a sleeve of a selected color over a capillary and move it to an easily visible position.

3.6 Rear of the instrument

In order to make transport easier, the instrument is equipped with rollers and a handle.

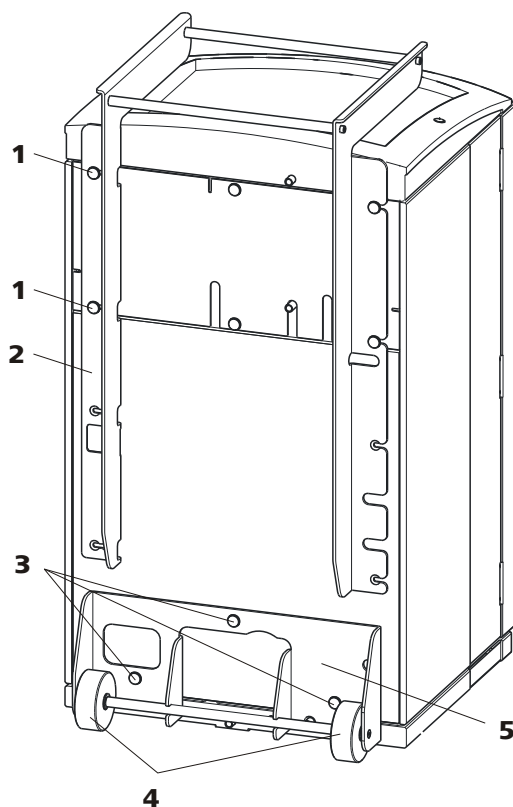


Figure 5 Rollers and handle

- | | |
|---|-------------------------|
| <p>1 Knurled screws</p> <p>For fastening the handle (5-2) and the rear panel of the detector chamber.</p> | <p>2 Handle</p> |
| <p>3 Knurled screws</p> <p>For fastening the roller holder (5-5).</p> | <p>4 Rollers</p> |
| <p>5 Roller holder</p> | |

Removing handle

- 1** Loosen knurled screws (5-**1**) and remove handle (5-**2**).

Removing rollers

Proceed as follows to remove the rollers:

- 1 Remove knurled screws (5-3).
- 2 Remove roller holder (5-5).

Mounting handle as MPak holder



- 1 Move handle (6-2) upwards and screw in the knurled screws (6-1) again.

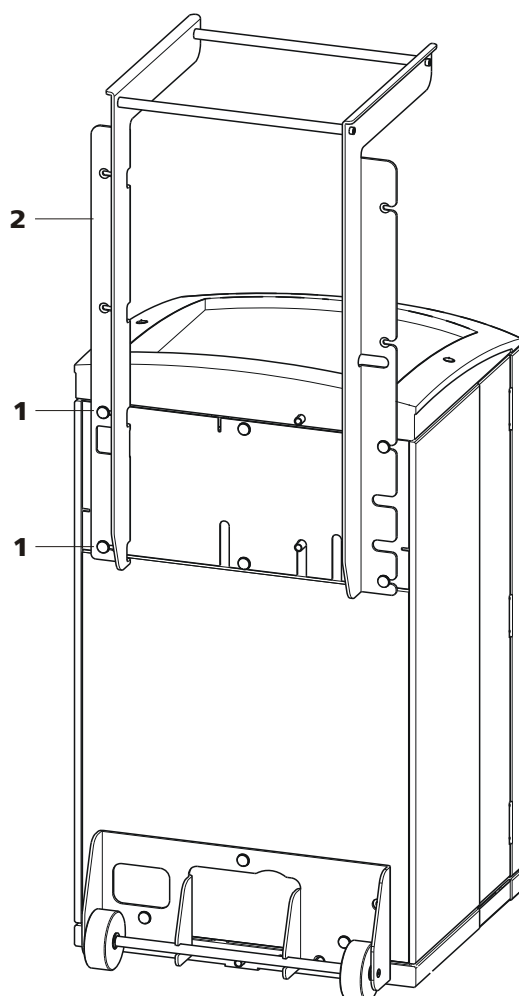


Figure 6 Handle as MPak holder

1 Knurled screws
For fastening the handle (6-2) and the rear panel of the detector chamber.

For fastening the handle (6-**2**) and the rear panel of the detector chamber.

2 Handle
Extended. As holder for MPaks (eluent bag).

Extended. As holder for MPaks (eluent bag).

Positioning and connecting the detector

The instrument is delivered without detector. For information on how to place and connect the detector, please refer to the manual of the detector.

Transport locking screws

To avoid damage to the high pressure pump and the vacuum pump during transport, the pumps are secured with transport locking screws.

Remove these transport locking screws before the initial start-up.

Removing transport locking screws

- 1** Remove all of the transport locking screws with the 4 mm hexagon key (6.2621.030) and keep them in a safe place.



WARNING

In order to avoid damage to the pumps, the transport locking screws must be remounted each time the instrument undergoes major transport.

Leak sensor

The leak sensor detects escaping liquid which collects in the base tray of the instrument.

To activate the leak sensor, the leak sensor connector plug (7-2) must be connected, the instrument switched on and the leak sensor switched to **active** in the software.

Connecting the leak sensor

- 1 Plug the leak sensor connector plug (7-2) into the leak sensor connection socket (7-1) on the rear of the instrument (see figure 7, page 23).



Figure 7 Connection of the leak sensor on the rear of the instrument

- | | |
|--|--|
| <p>1 Leak sensor connection socket
Is labeled with "Leak Sensor".</p> <p>3 Leak sensor connection cable
Is firmly mounted on the rear of the instrument.</p> | <p>2 Leak sensor connector plug</p> |
|--|--|

3.6.5 Drainage tubings

Liquid that escapes in the flask holder or in the detector chamber is directed through the drainage tubings into the base tray, past the leak sensor into the waste container. This ensures that any leaks in the system will be detected by the leak sensor.

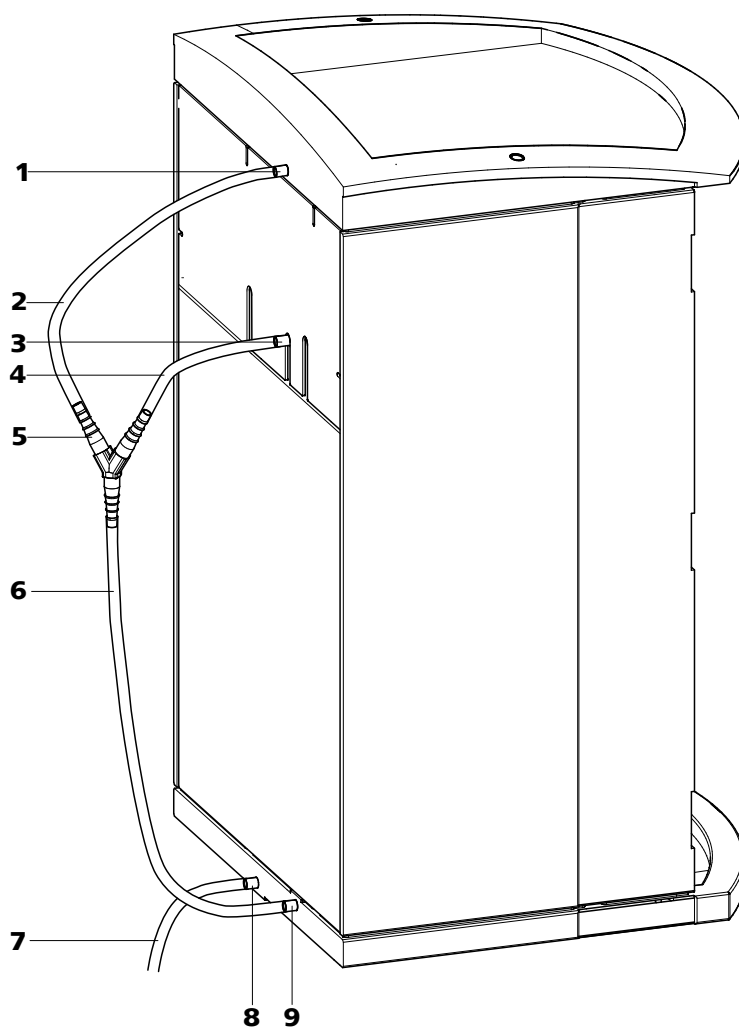


Figure 8 Drainage tubings

- | | |
|--|--|
| <p>1 Drainage tubing connector
For draining escaped liquid from the flask holder.</p> | <p>2 Drainage tubing
Section of the 6.1816.020 silicon tubing. For draining escaped liquid from the flask holder.</p> |
| <p>3 Drainage tubing connector
For draining escaped liquid from the detector chamber.</p> | <p>4 Drainage tubing
Section of the 6.1816.020 silicon tubing. For draining escaped liquid from the detector chamber.</p> |
| <p>5 Y connector 6.1807.010
For connecting the two drainage tubings (8-2) and (8-4).</p> | <p>6 Drainage tubing
Section of the 6.1816.020 silicon tubing. Guides escaped liquid to the leak sensor.</p> |

Section of the 6.1816.020 silicon tubing.
Guides escaped liquid into a waste container.

For draining the escaped liquid from the base tray through the connected drainage tubing.

For supplying escaped liquid through the connected drainage tubing to the leak sensor.

Proceed as follows to install the drainage tubings:

Installing drainage tubings

- 1 Plug drainage tubing (8-2) into the drainage tubing connector (8-1) of the flask holder and shorten to required length.
- 2 Plug drainage tubing (8-4) into the drainage tubing connector (8-3) of the detector chamber and shorten to required length.
- 3 Connect drainage tubing (8-2) from the flask holder and drainage tubing (8-4) from the detector chamber to the Y connector (8-5).
- 4 Connect drainage tubing (8-6) to the Y connector (8-5), shorten to required length and plug the other end of the drainage tubing into the drainage tubing connector (8-9) of the base tray.
- 5 Plug drainage tubing (8-7) into the drainage tubing connector (8-8) of the base tray and guide the other end into a waste container.

3.7 Capillary and cable feed-throughs

Several openings have been integrated for feeding through capillaries and cables. They are located on the doors (*see figure 9, page 26*), on the rear panel or below the flask holder or above the base tray (*see Figure 10, Page 27*).

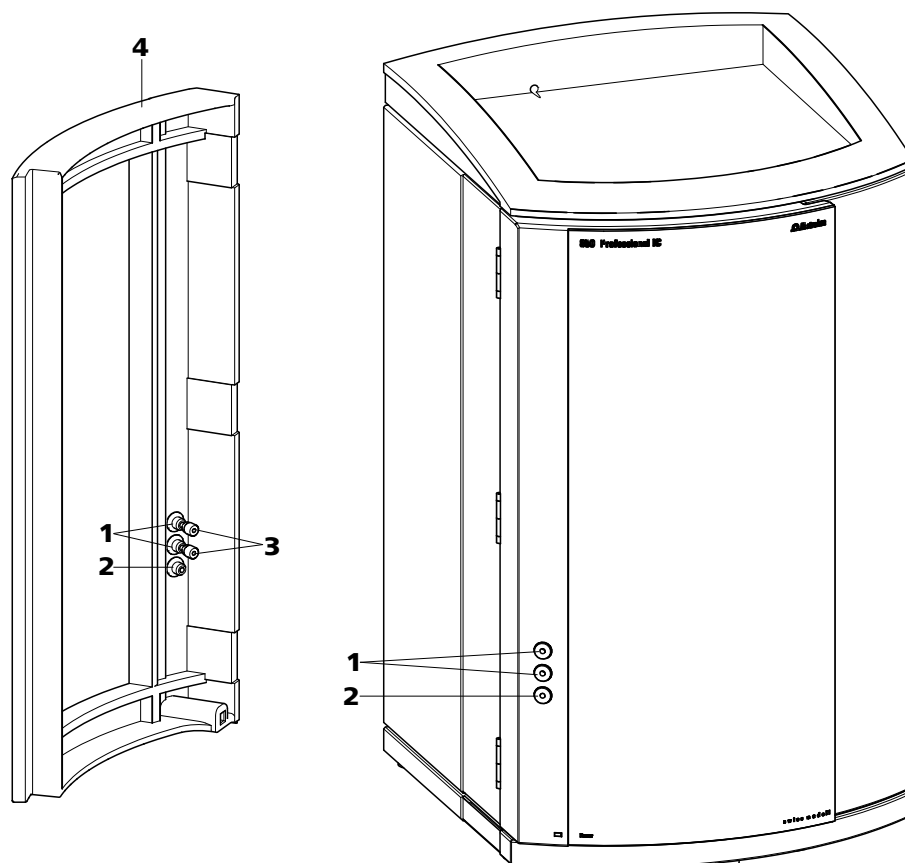


Figure 9 Capillary feed-throughs on the door

1 Luer connectors

For connecting a 6.2816.020 syringe. For manual sample feeding.

**3 PEEK pressure screws, short
6.2744.070**

2 Capillary feed-through

4 Door

Do not feed capillaries through the Luer connectors (9-**1**). The capillaries are fastened with PEEK pressure screws (9-**3**) from inside to the Luer connector. From outside, liquid can be aspirated or injected with a syringe.



1	Side panel (right) Right panel.	2	Rear of the instrument
3	Side panel (left) Left panel.	4	Front of the instrument
5	Capillary feed-through Upper. From front to right.	6	Capillary feed-through Upper. From front to back.
7	Capillary feed-through Upper. From front to back.	8	Capillary feed-through Upper. From front to left.
9	Capillary feed-through Lower. From front to right.	10	Capillary feed-through Lower. From front to back.
11	Capillary feed-through Lower. From front to back.	12	Capillary feed-through Lower. From front to left.

3.8 Eluent

3.8.1 Connecting the eluent bottle

The eluent is aspirated out of the eluent bottle via the eluent aspiration tubing (11-1).

The eluent aspiration tubing is connected to the eluent degasser (*see chapter 3.9, page 32*). The tubing must be threaded through a suitable capillary feed-through (*see chapter 3.7, page 25*) of the instrument before the other end can be equipped.

The following accessories are needed for equipping the eluent aspiration tubing:

- 6.1602.160 eluent bottle cap GL 45
- 6.2744.210 tubing adapter for aspiration filter
- 6.2821.090 aspiration filter

Proceed as follows for equipping the eluent aspiration tubing:

Assembling eluent aspiration tubing

- 1** Guide the free end of the eluent aspiration tubing (11-1) out of the instrument through a suitable capillary feed-through.
- 2 Installing the eluent bottle cap (6.1602.160)**
 - Slide tubing nipple (11-2) and O-ring (11-3) onto the eluent aspiration tubing (11-1).
 - Push eluent aspiration tubing (11-1) through the bottle cap (11-4) and screw tight.



3 Mounting the aspiration filter

- Insert the filter holder (12-**1**) into the aspiration filter (12-**2**) and screw it tightly.

850 Professional IC Anion MSM-HC HP-Gradient

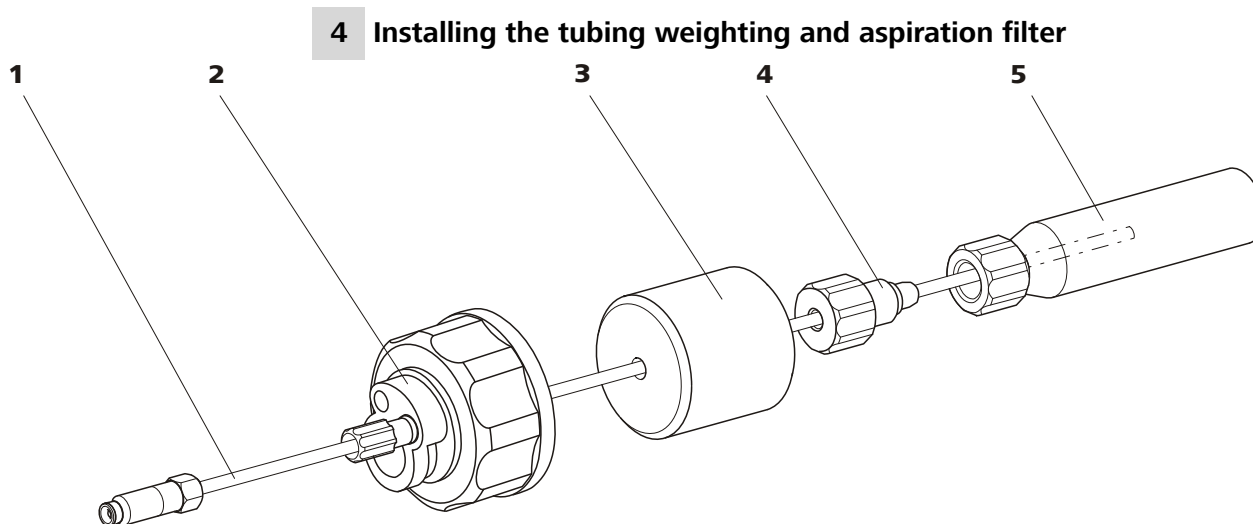


Figure 13 Installing the tubing weighting and aspiration filter

1	Eluent aspiration tubing (6.1834.080)	2	Eluent bottle cap (6.1602.160)
3	Tubing weighting From accessory set (6.2744.210).	4	Clamping screw From accessory set (6.2744.210).
5	Aspiration filter (6.2821.090) With filter holder from accessory set (6.2744.210).		

- Slide the tubing weighting (13-3) onto the eluent aspiration tubing (13-1).
- Slide the clamping screw (13-4) onto the eluent aspiration tubing (13-1).
- Insert eluent aspiration tubing (13-1) into the aspiration filter (13-5). The end of the tubing should reach approximately to the center of the aspiration filter.
- Screw the clamping screw (13-4) to the filter holder (12-1).

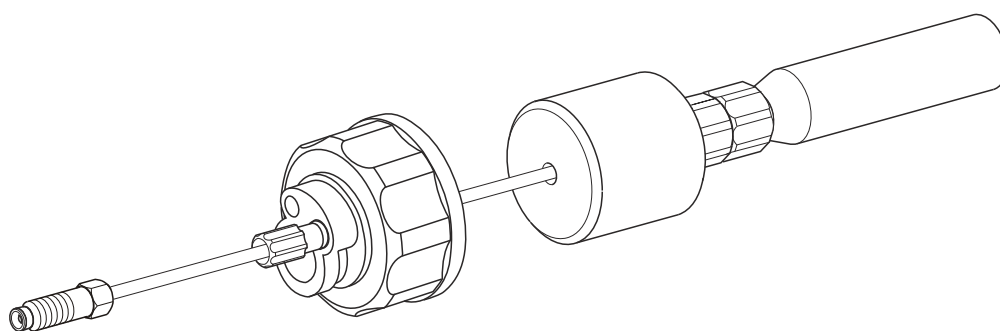


Figure 14 Eluent aspiration tubing fully equipped

5 Mounting the eluent aspiration tubing to the eluent bottle

- Insert the eluent aspiration tubing into the eluent bottle (**15-10**).

3	Cotton	4	CO₂ adsorber Adsorbs CO ₂ from the air (e.g. Merck soda lime with indicator, no. 6839.1000).
5	Eluent	6	Aspiration filter (6.2821.090)
7	Filter holder From accessory set (6.2744.210).	8	Clamping screw From accessory set (6.2744.210).
9	Tubing weighting From accessory set (6.2744.210).	10	Eluent bottle (6.1608.070)
11	Bottle cap (6.1602.160)	12	SGJ clip (6.2023.020)
13	Tubing nipple	14	Threaded stopper

3.9 Eluent degasser

Gas bubbles in the eluent lead to an unstable baseline, as high pressure pumps can transport liquids, but not gases. This is why the eluent must be degassed before it reaches the high-pressure pump.

The eluent degasser removes gas bubbles and dissolved gases from the eluent. For degassing, the eluent flows into a vacuum chamber through a special fluoropolymer capillary.



NOTICE

The eluent degasser is already installed in the newly delivered instrument. The following installation instructions must only be followed, if the connections to the degasser had to be disconnected for maintenance.

The diagram illustrates the Eluent Degasser assembly. It features two circular ports on the left, labeled 'In' (top) and 'Out' (bottom). The 'In' port is connected to a tube labeled '1', and the 'Out' port is connected to a tube labeled '2'. Both tubes lead to a central degasser unit, which is a cylindrical component with a flange on the left and a threaded cap on the right. The degasser unit is labeled '3' and '4'. The tubes are labeled '5' and '6'.

Figure 16 Eluent degasser

1	Eluent degasser input	2	Eluent degasser output
3	Tubing flare With tubing nipple.	4	Clamping screw
5	Eluent aspiration tubing (6.1834.080) For aspirating the eluent. The clamping screw (16-4) is firmly mounted.	6	Connection tubing (6.1834.090) Connection from the eluent degasser to the high-pressure pump (<i>see chapter 3.10, page 34</i>). The clamping screw (16-4) is firmly mounted.

CAUTION

The clamping screws (16-4) must be tightened carefully. Use the wrench (6.2621.050) to do this.

- Insert the eluent aspiration tubing (16-5) into the eluent degasser input (16-1).
- Carefully tighten the clamping screw (16-4).

- Insert connection tubing (16-6) (the end with the longer clamping screw (16-4)) into the eluent degasser output (16-2).
- Carefully tighten the clamping screw (16-4).
- Connect the other end of the connection capillary (16-6) (with the shorter clamping screw) to the high-pressure pump (17-9) (see "Connecting inlet to the high pressure pump", page 35).

3.10 High pressure pump

The intelligent and low pulsation high pressure pump pumps the eluent through the system. It is equipped with a chip on which its technical specifications and "life history" (operating hours, service data, ...) are saved.

The purge valve is used for deaerating (see chapter 3.10.2, page 36) the high pressure pump.

3.10.1 Capillary connections high pressure pump/purge valve



NOTICE

All of the capillary connections of the high pressure pump and the purge valve are already installed in the newly delivered instrument.

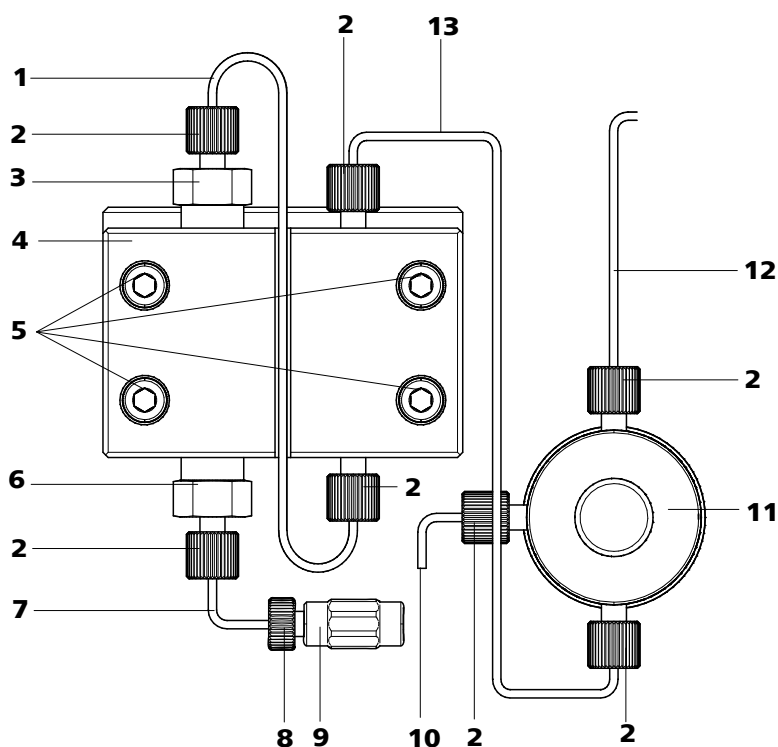


Figure 17 Capillary connections high pressure pump/purge valve

- 1 Connection capillary**
PEEK capillary, connects main piston and auxiliary piston.
- 3 Outlet valve holder**

- | | |
|---|--|
| 2 | PEEK pressure screw, short
(6.2744.070) |
| 4 | Pump head (6.2824.110) |

- 

The eluent aspiration capillary is already installed in the newly delivered instrument. The following installation instructions need **not** be carried out at the time of initial installation.

1 Pressure screw For connecting the coupling (18-2) to the pump head inlet capillary (17-7). Can be ordered together with the coupling under the number (6.2744.230).	2 Coupling (6.2744.230) For connecting the eluent connection capillary (18-4) to the input of the high pressure pump.
---	--



- ### 3 Clamping screw

- #### 4 Eluent aspiration tubing

Eluent aspiration tubing (6.1834.080) or (6.1834.090).

- ## 5 Backup ring

1 Connecting coupling

Fasten the coupling (18-2) with a pressure screw (18-1) on the pump head inlet capillary (17-7).

2 Connecting eluent aspiration tubing



CAUTION

The clamping screws must be tightened carefully. To tighten, grip the coupling (**18-2**) with the key (6.2739.000) and grip the clamping screw (**18-3**) with the wrench (6.2621.050).

- Plug the eluent aspiration tubing (18-4) into the coupling (18-2).
- Tighten the clamping screw (18-3).

3.10.2 Deaerating the high pressure pump

The high pressure pump will only operate perfectly if the pump head contains no more air bubbles. Therefore it must be deaerated during initial start-up and after every change of eluent.



CAUTION

The high pressure pump must **not** be deaerated before the initial start-up (see chapter 4.1, page 60).

Deaerate the high pressure pump as follows (see figure 19, page 37):

Deaerate the high pressure pump

The instrument must be connected to the PC and switched on to deaerate the high pressure pump.



5 Aspirating eluent

- Aspirate with the syringe (**19-1**) until bubble-free eluent flows into the syringe.

6 Completing deaerating

- Switch off high pressure pump.
- Close rotary knob (19-7).
- Remove syringe (19-1) from the Luer connector (19-2).
- Pull the purging needle (19-3) out of the deaerating capillary (19-4).

3.11 Inline filter

Between the purge valve and the pulsation absorber the inline filter (6.2821.120) is installed as protection against particles.

Inline filters protect the separation column securely against possible contamination from the eluent. Inline filters can however also just as well be used for the purpose of protecting the suppressor against contaminations in the regeneration or rinsing solutions. The filter platelets with a pore size of 2 µm can be replaced quickly and easily. They remove particles like e. g. bacteria and algae from the solutions.



NOTICE

The inline filter is already installed in the newly delivered instrument. The following installation instructions need **not** be carried out at the time of initial installation.

Installing the inline filter



CAUTION

Observe the flow direction marked on the filter housing for the connection of the inline filter.

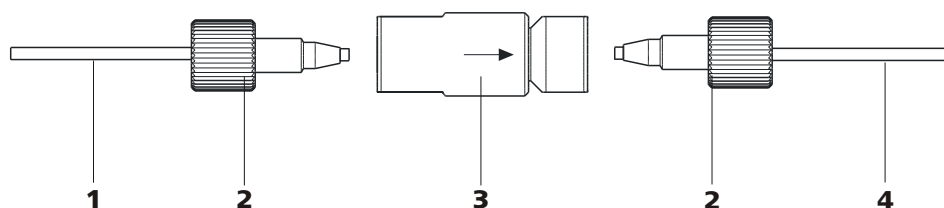


Figure 20 Connecting the inline filter

1 Connection capillary

Connects the purge valve with the inline filter

3 Inline filter (6.2821.120)

Protects against particles.

2 PEEK pressure screws, short (6.2744.070)

4 Connection capillary

Connects the inline filter with the pulsation absorber.

- 1** Screw on the connection capillary running from the purge valve to the input side of the inline filter using a pressure screw (6.2744.070).
- 2** Screw on the connection capillary running to the pulsation absorber to the output side of the inline filter using a pressure screw (6.2744.070).

3.12 Pulsation absorber



NOTICE

The pulsation absorber is already installed in the newly delivered instrument.



CAUTION

The pulsation absorber is maintenance-free and may not be opened.

The pulsation absorber protects the separation column from damage caused by pressure fluctuations when switching the injection valve, and reduces interfering pulsations during highly sensitive measurements. In order to ensure these functionalities, it must be connected between the high pressure pump (see chapter 3.10, page 34) and injection valve (see chapter 3.14, page 42).

The pulsation absorber can be operated in both directions.

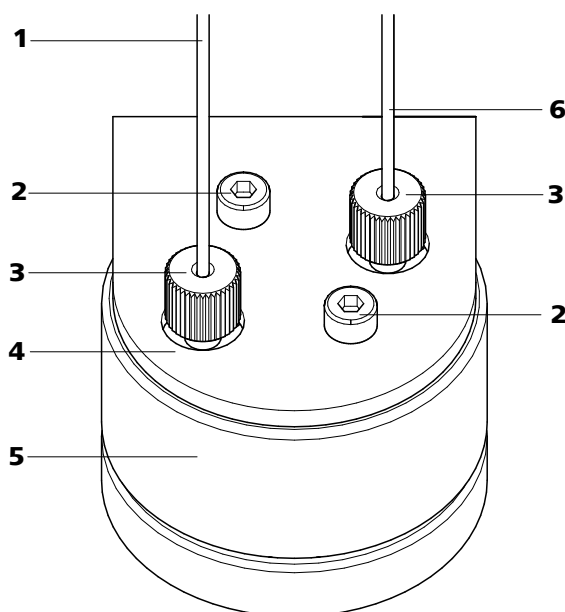


Figure 21 Pulsation absorber – Connection

1	Connection capillary Connection to the inline filter.	2	Fastening screws
3	PEEK pressure screws, short (6.2744.070)	4	Holder for pulsation absorber
5	Pulsation absorber (6.2620.150)	6	Connection capillary Connection to the injection valve.

3.13 Sample degasser

The sample degasser removes gas bubbles and dissolved gases from the sample. For degassing, the sample flows into a vacuum chamber through a special fluoropolymer capillary.

Gas bubbles in the sample lead to poor reproducibility, as the amount of sample in the sample loop would not always be the same. This is why samples (containing gas) should be degassed prior to injection. The sample is drawn in through a degassing chamber prior to injection and any gas bubbles are automatically removed.



NOTICE

The rinsing time increases by at least 2 minutes when the sample degasser is used.



Connecting the sample degasser

- 1 Remove and keep the threaded stoppers (6.2744.220) from the inlet and outlet of the sample degasser.
- 2 Connect the end of the sample aspiration capillary (6.1803.040) connected to the injection valve to the output of the sample degasser (22-**2**), using a long PEEK pressure screw (22-**3**).
- 3 Connect the connection capillary (6.1803.040) to the input of the sample degasser (22-**1**), using a long PEEK pressure screw (22-**3**).
- 4 Guide the other end of the connection capillary out of the instrument through a capillary feed-through and connect it with the Sample Processor, if applicable.



If the sample degasser is not used, the input and output **must** be sealed with the threaded stoppers (6.2744.220).



NOTICE

The column input capillary is already threaded into the capillary recesses of the column thermostat at the time the instrument is first delivered. The following installation instructions need **not** be carried out at the time of initial installation.

Threading capillaries

- 1 Introduce column input capillary into the column thermostat via a suitable capillary feed-through (25-1).
- 2 Slide the column input capillary from below into the outer of the two capillary recesses (25-2). Slide it through under the holder plate until it emerges again at the top.
- 3 Carefully bend the column input capillary downward and slide from from above to below through the inner capillary recess until it emerges at the lower edge of the holder plate.



NOTICE

The columns (guard column and separation column) may only be installed after the initial start-up (see chapter 4.1, page 60).

- **Before initial start-up:**
Fasten the 6.2744.040 coupling with a 6.2744.010 pressure screw to the end of the column input capillary.
- **After initial start-up:**
Fasten the guard column (if used) or the separation column with a 6.2744.010 pressure screw to the end of the column input capillary.

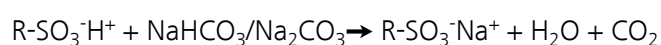
3.16 High Capacity Metrohm Suppressor Module (MSM-HC)

3.16.1 General information on the MSM-HC

The MSM-HC is used for chemical suppression during anion analysis if eluents with a high ionic concentration are used or if the concentration is being significantly increased in the course of a gradient application. It consists of 3 suppressor units in total, which are used for suppression, regenerated with sulfuric acid or rinsed with ultra pure water in rotation.

Suppression reaction in the MSM-HC

When using a carbonate eluent, the following reaction (amongst others) occurs in the MSM-HC:



3.16.2 Connecting the MSM-HC



CAUTION

To protect the MSM-HC against foreign particles or bacterial growth, a 6.2744.180 pump tubing connection with filter must be mounted between the peristaltic pump and the inlet capillaries of the MSM-HC.

The three inputs and outputs of the suppressor units numbered with 1, 2 and 3 on the MSM-HC each have 2 fixed mounted PTFE capillaries (see *figure 26, page 48*).

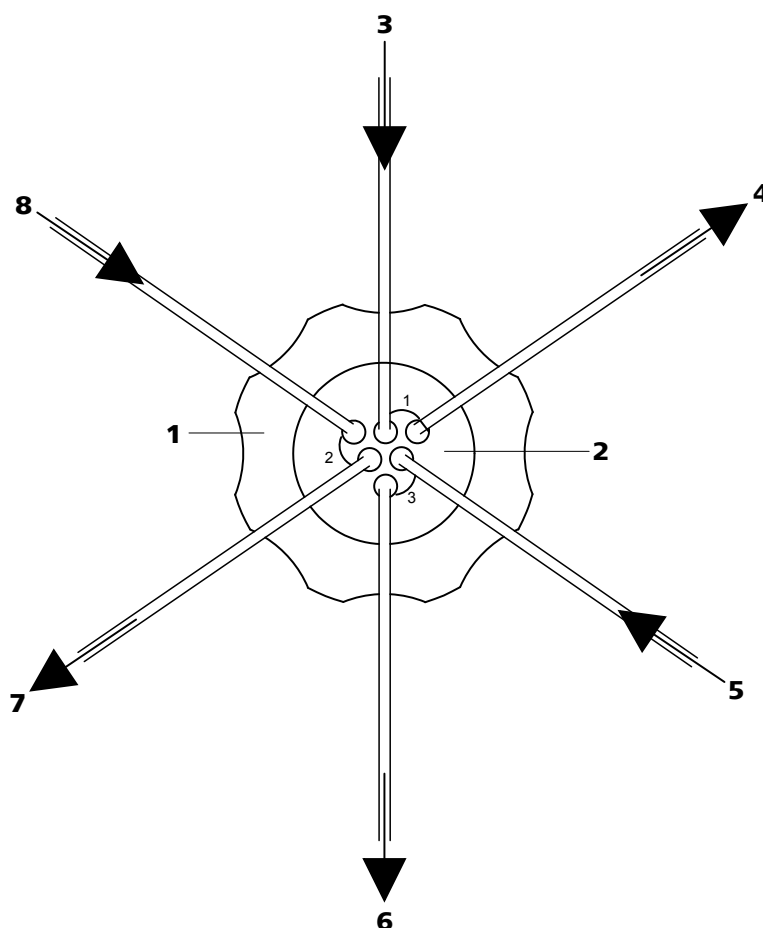


Figure 26 MSM-HC – Connectors

1	Union nut	2	MSM-HC connecting piece 6.2835.010
3	Eluent inlet capillary Labeled with <i>in.</i>	4	Eluent outlet capillary Labeled with <i>out.</i>
5	Rinsing solution inlet capillary Labeled with <i>rinsing solution.</i>	6	Rinsing solution outlet capillary Labeled with <i>waste rins..</i>
7	Regeneration solution outlet capillary Labeled with <i>waste reg..</i>	8	Regeneration solution inlet capillary Labeled with <i>regenerant.</i>

The PTFE capillaries firmly mounted on the MSM-HC are connected to the other components of the IC system as follows:



CAUTION

As the PTFE capillaries are very soft, the pressure screws should not be overtightened.

Flattened capillaries can be shortened with the help of a capillary cutter.

1 Connecting the eluent inlet capillary

- ## 2 Connecting the eluent outlet capillary



OR

3 Connecting the rinsing solution inlet capillary

- #### 4 Connecting the rinsing the solution outlet capillary

- ## 5 Connecting the regeneration solution inlet capillary

- ## 6 Connecting the regeneration solution outlet capillary

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1	PEEK pressure screws, short (6.2744.070)	2	Tubing olive (6.2744.034)
3	Stopper The colors of the stopper indicate the inner diameter of the pump tubing.	4	Tubing cartridge (6.2755.000)
5	Contact pressure lever	6	Union nut
7	Adapter	8	Tubing olive Either with filter holder (6.2744.180) or without filter holder (6.2744.160).
9	Pump tubing (6.1826.xx0)	10	Snap-action lever

1 Removing the tubing cartridge

2 Connecting the aspiration side

■■■■■■■ 51

3 Connecting the pressure side



NOTICE

Depending on the use of the peristaltic pump, on the pressure side you can either connect:

- **Case A:** a 6.2744.180 pump tubing connection **with filter**
(see figure 29, page 52) or
- **Case B:** a 6.2744.160 pump tubing connection **without filter**
(see figure 30, page 53).

For pumping the auxiliary solutions to the MSM or to the SPM, a 6.2744.180 pump tubing connection **with filter must** be used.

Case A: 6.2744.180 pump tubing connection with filter:

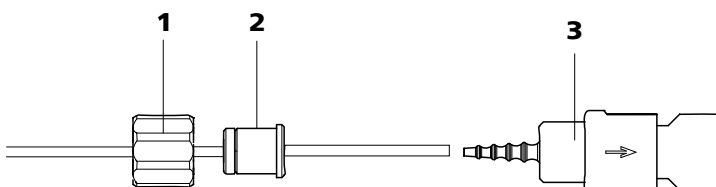


Figure 29 Install pump tubing connection with filter

- ## 1 Union nut

- ## 2 Adapter

- ### 3 Tubing olive with filter holder

- Slide union nut (29-**1**) onto the pump tubing.
- Select a suitable adapter (29-**2**) and slide it onto the pump tubing. The type of adapter depends on the pump tubing (*see table 1, page 53*).
- Place the tubing olive with filter holder (29-**3**) onto the pump tubing.
- Screw the union nut (29-**1**) onto the tubing olive (29-**3**).

or

Case B: 6.2744.160 pump tubing connection without filter:



2 Adapter

3 Tubing olive

- Slide union nut (30-1) onto the pump tubing.
- Select a suitable adapter (30-2) and slide it onto the pump tubing. The type of adapter depends on the pump tubing (*see table 1, page 53*).
- Place the tubing olive (30-3) onto the pump tubing.
- Screw the union nut (30-1) onto the tubing olive (30-3).

4 Inserting the pump tubing

- #### 4 Inserting the pump tubing
- Press the contact pressure lever all the way down.
 - Place the pump tubing in the tubing cartridge. The stoppers (28-3) must snap into the corresponding holders of the tubing cartridge.

5 Inserting the tubing cartridge

- ## 5 Inserting the tubing cartridge
- Hang the tubing cartridge in the mounting pin and press in the cartridge holder until the snap-action lever snaps in.

6 Connecting the capillaries

- ## 6 Connecting the capillaries
- Screw the respective capillaries tightly to the two tubing olives with PEEK pressure screws (28-1).

Table 1 Pump tubings and suitable adapters

Pump tubing	Adapter
6.1826.020 (blue/blue)	
6.1826.310 (orange/green)	
6.1826.320 (orange/yellow)	
6.1826.330 (orange/white)	

3.18 Connecting the instrument to a computer



NOTICE

If the instrument is connected to the computer, then it must be switched off.

Accessories

For this step, you need the following accessories:

- USB connecting cable (6.2151.020)

Connecting the USB cable

- 1 Insert the USB cable into the computer connection socket on the rear of the instrument labeled *PC*.
- 2 Insert the other end into a USB port on the computer.

3.19 Connecting the instrument to the power grid



WARNING

Electric shock from electrical potential

Risk of injury by touching live components or through moisture on live parts.

- Never open the housing of the instrument while the power cord is still connected.
- Protect live parts (e.g. power supply unit, power cord, connection sockets) against moisture.
- Unplug the power plug immediately if you suspect that moisture has gotten inside the instrument.
- Only personnel who have been issued Metrohm qualifications may perform service and repair work on electrical and electronic parts.

Connecting the power cord

Accessories

Power cord with the following specifications:

- Length: max. 2 m
- Number of cores: 3, with protective conductor

- Instrument plug: IEC 60320 type C13
- Conductor cross-section 3x min. 0.75 mm² / 18 AWG
- Power plug:
 - according to customer requirement (6.2122.XX0)
 - min. 10 A



NOTICE

Do not use a not permitted power cord!

1 Plugging in the power cord

- Plug the power cord into the instrument's power socket.
- Connect the power cord to the power grid.

3.20 Guard column

The use of guard columns serves for protecting the separation columns and increasing their service life considerably. The guard columns available from Metrohm represent either actual guard columns or are so-called guard column cartridges which are used together with a cartridge holder. The installation of a guard column cartridge in the associated holder is described in the leaflet of the guard columns.



NOTICE

Information regarding which guard column is suitable for your separation column can be found in the **Metrohm IC Column Program** (which is available from your Metrohm agent), the leaflet provided along with your separation column, the product information on the separation column at <http://www.metrohm.com> (product area Ion Chromatography), or obtained directly from your agent.



CAUTION

New guard columns are filled with solution and are sealed on both sides with stoppers or caps, respectively. Before using the guard column, you need to ensure that this solution is miscible with the eluents used (observe manufacturer's data).



NOTICE

Metrohm recommends, always to work with guard columns. These protect the separation column and can be exchanged regularly.

Connecting and rinsing the guard column

1 Connecting the guard column



When inserting the guard column, always ensure that it is inserted correctly corresponding to the flow direction (if indicated).

- Remove sealing caps and/or stoppers from the guard column.
- Fasten the input of the guard column to the column inlet capillary using a short PEEK pressure screw (6.2744.070).
- In case the guard column is mounted on the separation column with a connecting capillary, connect the included connection capillary, which is included with the guard column, to the output of the guard column using the PEEK pressure screw, which is also included.

2 Rinsing the guard column

- Place beaker under the outlet capillary of the guard column.
- Set the flow rate of the high pressure pump according to the data given in the leaflet of the separation column.
- Start the high pressure pump and rinse the guard column approx. 5 minutes with eluent.
- Switch off the high pressure pump again.

3.21 Separation column

The intelligent separation column (iColumn) is the heart of the ion chromatographic analysis. It separates the different components corresponding to their interactions with the column. Metrohm separation columns are equipped with a chip on which their technical specifications and their history (first use / setting up, operating hours, injections, ...) are saved.



NOTICE

Information regarding which separation column is suitable for your application can be found in the **Metrohm IC Column Program**, the product information for your separation column at <http://www.metrohm.com> in the product area ion chromatography, or obtained directly from your agent.



CAUTION

New separation columns are filled with solution and are sealed on both sides with stoppers. Before using the column, you need to ensure that this solution is miscible with the eluents used (observe manufacturer's data).

You can find the separation columns and guard columns currently available from Metrohm in the Metrohm IC Column Program, or in the Internet at <http://www.metrohm.com> in the product area Ion Chromatography. A test chromatogram and an leaflet are provided along with each column. You can request detailed information on special IC applications in the corresponding "**Application Bulletins**" or "**Application Notes**", available in the Internet at <http://www.metrohm.com> in the Applications area or via the Metrohm agent responsible free of charge.



NOTICE

The separation column may only be installed after the **initial start-up** (see chapter 4.1, page 60) of the instrument. Until then, use the coupling (6.2744.040) instead of the guard and separation column.

1 Connect the separation column



- Remove stoppers from the separation column.
- Attach the guard column to the input of the separation column.
OR
Connect the input of the separation column to the outlet capillary of the guard column, using the PEEK pressure screw (6.2744.070) included.
OR
If no guard column is used (not recommended), connect the column input capillary to the input of the separation column, using a PEEK pressure screw (6.2744.070).

- Place beaker under the outlet end of the separation column.
- Set the flow rate of the high pressure pump according to the data given in the leaflet of the separation column.
- Start the high pressure pump and rinse the separation column approx. 10 minutes with eluent.
- Switch off the high pressure pump again.

- Fasten the column output capillary to the output of the separation column using a PEEK pressure screw (6.2744.070).
- Hang separation column with chip into the column holder.



The iColumns are equipped with a chip on which their operating data is saved. The chip has to be hooked into the chip holder provided for this so that the column recognition can function.

4 Start-up

The chapter *Start-up* is divided into 2 sections:

Initial start-up

The **initial start-up** is carried out during the **initial installation**.

Conditioning

Conditioning is carried out as a final installation step and each time after the system is started.

4.1 Initial start-up

The initial start-up is carried out during the initial installation. The entire system is rinsed before guard column and separation column are installed.



CAUTION

The separation column and guard column may not be installed for the initial start-up.

Make sure that the coupling (6.2744.040) is being used instead of the columns.

Perform the following steps during the initial start-up:

1 Preparing the software

- Start the PC program **MagIC Net™**.
- Open the **Equilibration** tab in MagIC Net™.
- Select (or create) a suitable method.

2 Preparing the instrument

- Ensure that the eluent aspiration tubing is immersed in the eluent and that there is enough eluent in the eluent bottle.
- Ensure that the aspiration tubings for the auxiliary solutions (regeneration solution and rinsing solution) are immersed into the respective solutions and that there is enough solution in the bottles.
- Switch on the instrument.

3 Starting equilibration

- In MagIC Net™, start the equilibration.

4 Deaerate the high pressure pump

- Deaerate the high pressure pump(s) via the purge valve (*see chapter 3.10.2, page 36*).

5 Set the contact pressure of the peristaltic pump**NOTICE**

This work step needs to be performed only if a peristaltic pump is being used.

- If peristaltic pumps are used, set the contact pressure (*see "Set flow rate", page 54*).

6 Rinsing the instrument without columns

- Rinse the instrument (without columns) with eluent for 5 minutes.

The instrument is now ready for the installation of the columns (*see chapter 3.20, page 56*).

4.2 Conditioning

After the installation and after switching on the instrument, the system must be conditioned with eluent until a stable baseline is reached.

**NOTICE**

After a change of eluent (*see chapter 5.4.2.3, page 66*), the conditioning time can lengthen considerably.

Conditioning the system

1 Preparing the software**CAUTION**

Ensure that the flow set is not higher than the flow permissible for the corresponding column (see column leaflet and chip data set).

- Start the PC program **MagIC Net™**.
- Open the **Equilibration** tab in MagIC Net™.



- Select (or create) a suitable method.

2 Preparing the instrument

- Ensure that the column is correctly mounted according to the flow direction indicated on the label (arrow must point in the direction of flow).
- Ensure that the eluent aspiration tubing is immersed in the eluent and that there is enough eluent in the eluent bottle.
- Ensure that the aspiration tubings for the auxiliary solutions (regeneration solution and rinsing solution) are immersed into the respective solutions and that there is enough solution in the bottles.

3 Checking leak-tightness

- In MagIC Net™, start the equilibration.
- Check all capillaries and their connections from the high pressure pump to the detector for signs of liquid escaping. If eluent escapes anywhere, tighten the corresponding pressure screw or loosen the connection, check the end of the capillary, shorten it with a capillary cutter if necessary, and restore the connection.

4 Conditioning the system

Rinse the system with eluent until the required stability of the baseline is attained (normally 30 minutes).

During this time, step the MSM to the next position every 10 minutes.

The instrument is now ready for measuring samples.

5.1 General notes

5.1.1 Care



The instrument requires appropriate care. Excess contamination of the instrument may result in functional disruptions and a reduction in the service life of the sturdy mechanics and electronics.



On the rear of the instrument, the drainage tubings must be mounted and the leak sensor must be plugged in and activated as protection against escaping liquids.

Spillages of chemicals and solvents should be cleaned up immediately. In particular, the plug connections on the rear panel of the instrument (especially the mains plug) should be protected from contamination.

5.1.2 Maintenance by Metrohm Service

Maintenance of the instrument is best carried out as part of an annual service, which is performed by specialist personnel from Metrohm. If working frequently with caustic and corrosive chemicals, a shorter maintenance interval is recommended. The Metrohm service department offers every form of technical advice for maintenance and service of all Metrohm instruments.

In order to avoid disturbing temperature influences, the entire system including the eluent bottle must be protected against direct sunlight.

If the instrument is not used for a longer period, the whole IC system (except the columns) must be rinsed salt free with methanol/ultrapure water (1:4), in order to prevent eluent salts from forming crystals which may cause subsequent damage.

5.3 Door



CAUTION

The door is made of PMMA (polymethylmetacrylate). It must never be cleaned with abrasive media or solvents.



CAUTION

Never use the door as a handle.

5.4 Eluent

5.4.1 Production

Chemicals used for manufacturing eluents must have a purity grade of at least "p.a.". Only ultrapure water (resistance $> 18.2 \text{ M}\Omega \cdot \text{cm}$) may be used for dilution (this generally applies for reagents which are used in ion chromatography).

Newly manufactured eluents always have to be microfiltered (0.45 μm filter).

The composition of the eluent plays a critical role in chromatographic analysis:

Concentration	An increase in the concentration generally leads to shorter retention times and faster separation, but also to a higher background conductivity signal.
pH	pH changes lead to shifts in dissociation equilibria and thus to changes in retention times.
Organic solvents	Adding an organic solvent (e.g. methanol, acetone or acetonitrile) to watery eluents generally speeds up lipophilic ions.



5.4.2 Operation

5.4.2.1 Supply bottle

The supply bottle with the eluent must be connected as indicated in *chapter 3.8.1, page 28*. This is particularly important for eluents with volatile solvents (e.g. acetone).

Condensation in the eluent bottle must be avoided. Drop formation can change the concentration ratio in the eluent.

In the case of very sensitive measurements, we recommend that the eluent be stirred constantly with a magnetic stirrer (e. g. the 2.801.0010 with 6.2070.000).

5.4.2.2 Aspiration filter

To protect the IC system from foreign particles, we recommend aspirating the eluent via an aspiration filter (6.2821.090) (*12-2*). This aspiration filter must be replaced if it takes on a yellowish discoloration, and at least every 3 months.

5.4.2.3 Changing the eluent

Prevent any precipitations when changing the eluent. Immediately successive solutions must always be miscible. If the system has to be rinsed with organic solvents, use several solvents with rising or falling lipophilicity.

5.5 High pressure pump

5.5.1 Protection



CAUTION

The pump head is filled ex works with methanol/ultrapure water. It must be ensured that the eluent used is freely miscible with the solvent remaining in the pump head.

To protect the high pressure pump against **foreign particles**, we recommend that the eluent undergoes a **microfiltration** (filter 0.45 µm) before being aspirated via a 6.2821.090 aspiration filter (*see "Assembling eluent aspiration tubing", page 28*).

Salt crystals between the piston and seal cause abrasion particles which can find their way into the eluent. These lead to contaminated valves, a rise in pressure and in extreme cases scratched pistons. It is therefore essential to ensure that **no precipitates** can occur (*see chapter 5.4.2.3, page 66*).

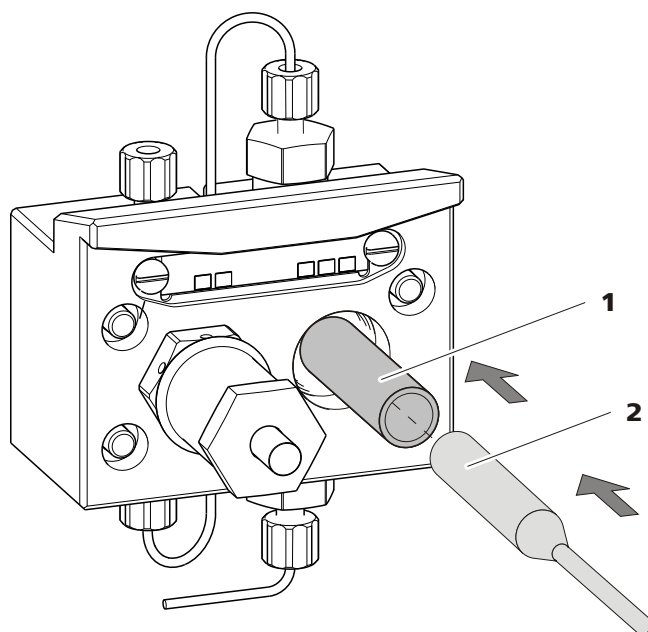


Figure 36 Inserting the piston seal into the pump head

4 Replacing the piston cartridge

Screw the assembled piston cartridge back into the pump head and tighten, first by hand, then additionally by approx. 15° with a wrench.

Cleaning the inlet valve and outlet valve

1 Removing valves

- Unscrew the connection capillary for the auxiliary piston (17-1) from the outlet valve holder.
- Unscrew the holders for the inlet and outlet valves and remove the valves (37-3) and (37-2).



2 Cleaning undissected valve

- Rinse the valve in eluent flow and counterflow direction using a spray bottle filled with ultrapure water, RBS solution or acetone.
- The rinsing effect is further increased through a short treatment (lasting for a maximum of 20 s) in an ultrasonic bath.



Only if this cleaning is useless, dismantle the valves separately and clean the components.

Dismantle every valve separately.



■■■■■■■ 73

- Place the valve with the seal faced downwards above the recess in the holder.
- Push the valve components out of the valve housing using the needle of the tool.

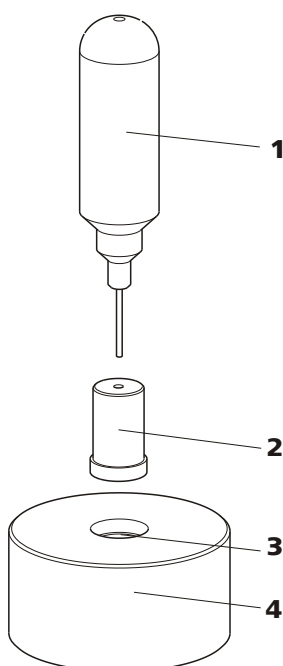


Figure 38 Dismantling valve

1 Needle For pushing the valve components out of the valve housing.	2 valve
3 Recess For collecting the valve components.	4 Holder

The components of the valve are collected in the recess of the holder.



NOTICE

The components of the valve are very small. In order not to lose them, put the components into a dish.

- The inlet valve and the outlet valve consist of the same, just differently arranged components (*see figure 39, page 75*).



- Place over the valve housing and hold it tightly.
- By tilting the tool, the valve components slide into the valve housing.
- Press the seal by hand well on the valve housing.

6 Checking the flow direction

Rinse the valve in the direction of the arrow on the valve housing and check whether liquid is escaping on the other end.

If this is not the case, the valve has to be dismantled again and be reassembled correctly (*see figure 39, page 75*).

7 Inserting the valves back into the pump head



CAUTION

If by mistake, the inlet valve is mounted instead of the outlet valve, an extreme pressure builds up within the working cylinder, which can destroy the piston seal!

When inserting the valves, please take into account that the liquid is being pumped through the pump head from bottom to top.

- Insert the inlet valve into the inlet valve holder the way the seal is visible.
- Screw the inlet valve holder into the bottom of the pump head and tighten with a wrench (37-4).
- Insert the outlet valve into the outlet valve holder the way the seal is visible.
- Screw the outlet valve holder into the top of the pump head and tighten with a wrench (37-1).

5.7 Inline sample preparation

To protect the separation column (*see chapter 3.21, page 58*) against foreign particles which can affect the separating efficiency, we recommend that all samples undergo a microfiltration (filter 0.45 µm). The ultrafiltration cell can be used for **filtration** (*see manual of the IC Equipment for Ultrafiltration*).

Samples containing significant amounts of **gas** should be degassed. The sample degasser (*see chapter 3.13, page 40*) is used for degassing.

Matrix-loaded samples (e.g. blood, oil) should be prepared for the measurement by means of dialysis (*see manual on the IC Equipment for Dialysis*).

If the concentration of the sample is too high, the sample should be **diluted** before feeding (*see documentation on the IC Equipment for Sample Dilution*).

For the sample preparation methods **Neutralization** (replacement of e.g. Na⁺ with H⁺) and **cation exchange** (replacement of e.g. heavy metals with H⁺), a sample preparation module (SPM) is used.

For an overview of all Metrohm inline sample preparation methods go to the following website: <http://misp.metrohm.com>

5.8 Rinsing the sample path

Before a new sample can be measured, the sample path must be rinsed with it so that the measuring result is not falsified by the previous sample (**Sample carry-over**).

In the case of automated sample feeding, the rinsing time should be at least 3 times the **transfer time**.

The transfer time is the time required by the sample to flow from the sample vessel to the end of the sample loop. The transfer time depends on the pump capacity of the peristaltic pump or the Dosino used, the total capillary volume and the volume of the gas removed by the sample degasser (if used) - in other words the amount of gas in the sample.

Ascertaining the transfer time

To ascertain the transfer time, proceed as follows:

1 Emptying the sample path

Pump air through the sample path (pump tubing, tubing connections, capillary in the degasser, sample loop) for several minutes until all liquid is displaced by the air.

2 Aspirating the sample and measuring time

Aspirate a sample typical for the later application and use a stop watch to measure the time required by the sample to travel from the sample vessel to the end of the sample loop.

The time measured corresponds to the "transfer time". The rinsing time should be at least 3 times the transfer time.

Checking the rinsing time

It is possible to determine whether the rinsing time is adequate via a direct measurement of the sample carry-over. Proceed as follows:

1 Preparing two samples

- **Sample A:** A typical sample for the application.
- **Sample sample B:** Ultrapure water.

2 Determining "Sample A"

Let "Sample A" pass through the sample path for the duration of the rinsing time, then inject and measure.

3 Determining "Sample B"

Let "Sample B" pass through the sample path for the duration of the rinsing time, then inject and measure.

4 Calculating the sample carry-over

The degree of the sample carry-over corresponds to the ratio of the peak areas of the measurement for sample B to the measurement for sample A. The lower the ratio, the lower the sample carry-over. This ratio can be modified by varying the rinsing time – thus allowing the rinsing time required for the application to be ascertained.

5.9.1 Operation



5.10.1 Protection

5.11.1 Protection

5.11.2 Operation



2 Regenerating the MSM-HC



CAUTION

The pump tubings made of PVC must not be used for rinsing with solutions containing organic solvents. In this case, other pump tubings must be used for rinsing.



NOTICE

The high pressure pump can be used for regeneration. For this, remove the guard column and separation column and connect the capillary directly to the MSM-HC (regenerate in opposite direction).

- Rinse the 3 suppressor units with the following solutions for about 15 minutes each:
 - **Contamination with heavy metals:**
1 mol/L H_2SO_4 + 0.1 mol/L oxalic acid
 - **Contamination with organic cationic complexing agents:**
0.1 mol/L H_2SO_4 / 0.1 mol/L oxalic acid / acetone 5%
 - **Severe contamination with organic substances:**
0.2 mol/L H_2SO_4 / acetone $\geq$ 20%

3 Connecting the MSM-HC to the IC system

- Reconnect the MSM-HC to the IC system. If the capacity problems remain, the MSM-HC rotor A must be replaced (*see chapter 5.11.3.3, page 86*).

5.11.3.2 Cleaning the MSM-HC

It may be necessary to clean the MSM-HC in the following cases:

- Increased backpressure onto the connection tubings of the MSM-HC.
- Blockage of the MSM-HC which cannot be eliminated (solutions can no longer be pumped through the MSM-HC).
- Jamming of the MSM-HC which cannot be eliminated (MSM-HC can no longer be switched over).

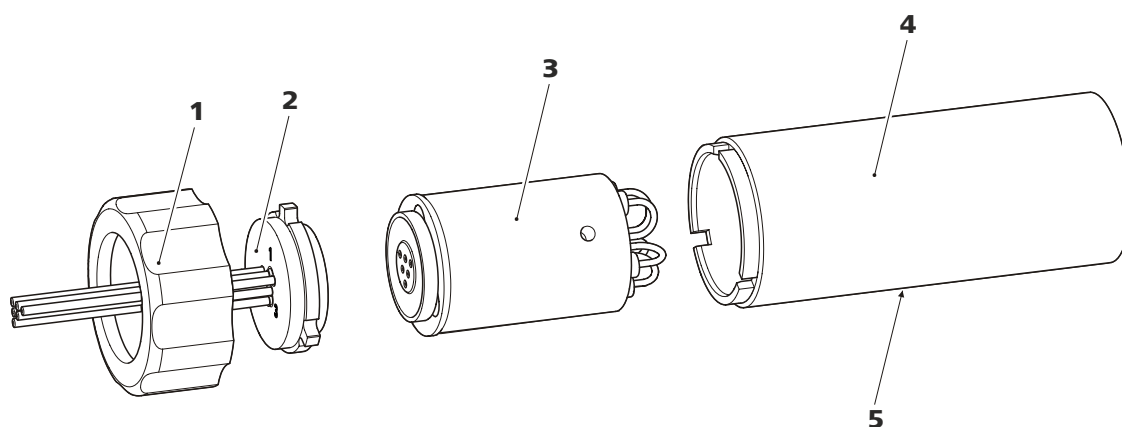


Figure 41 MSM-HC – Components

1 Union nut**3 MSM-HC rotor A 6.2842.000****5 Slot in the MSM-HC housing****2 MSM-HC connecting piece 6.2835.010****4 MSM-HC housing**

Cleaning the MSM-HC

Clean the MSM-HC as follows:

1 Disconnecting the MSM-HC from the IC system

- Switch off the instrument.
- Disconnect the MSM-HC from the separation column, peristaltic pump and detector.

2 Dismantling the MSM-HC

- Unscrew union nut (41-1) from the MSM-HC housing (41-4).
- Pull the MSM-HC connecting piece (41-2) and the MSM-HC rotor A (41-3) out of the MSM-HC housing (41-4). The connecting piece and the MSM-HC rotor A normally stick to one another – if this is not the case: Take a sharp object, insert into the slot (41-5) in the MSM-HC housing, and pull out the MSM-HC rotor A (41-3) in this way.
- Detach the MSM-HC connecting piece (41-2) from MSM-HC rotor A (41-3).

3 Cleaning the supply and discharge lines

- Connect in turn each of the 6 capillary tubings fastened on the MSM-HC connecting piece (41-2) on the high pressure pump (see chapter 3.10, page 34) and pump through ultra pure water.

- Check whether solution emerges at the MSM-HC connecting piece (41-2). If one of the supply or discharge lines remained blocked, the MSM-HC connecting piece (41-2) must be replaced (order number 6.2835.010).

4 Cleaning the MSM-HC rotor A

- Clean the sealing surface of the MSM-HC rotor A (41-3) with ethanol using a lint-free cloth.

5 Inserting the MSM-HC rotor A



CAUTION

An incorrectly inserted MSM rotor A (41-3) can be **destroyed** during start-up.

- Insert the MSM-HC rotor A (41-3) into the MSM-HC housing (41-4) in such a way that the tubing connections on the rear of MSM-HC rotor A fit into the corresponding recesses inside the MSM-HC housing and one of the three holes of the MSM-HC rotor A is visible from below in the slot (41-5) of the MSM-HC housing.
- If the MSM-HC rotor A (41-3) is correctly inserted, its sealing surface will be approx. 4 mm within the MSM-HC housing (41-4). If this is not the case, the MSM-HC rotor A must be moved into the right position from below using a sharp object (e.g. screwdriver).

6 Cleaning the MSM-HC connecting piece

- Clean the sealing surface of the MSM-HC connecting piece (41-2) with ethanol using a lint-free cloth.

7 Inserting the MSM-HC connecting piece

- Insert the MSM-HC connecting piece (41-2) into the MSM-HC housing (41-4) in such a way that the connector 1 is on top and the three pins of the connecting piece fit into the corresponding recesses on the MSM-HC housing (41-4).
- Reattach the union nut (41-1) onto the coupling and tighten.

8 Connecting and conditioning the MSM-HC

- Reconnect the MSM-HC to the IC system.
- Before switching the MSM-HC over for the first time, rinse the three suppressor units with solution for 5 minutes.

Order number	Name	Material	Inner diameter	Use
6.1826.340	Pump tubing LFL (black/black), 3 stoppers	PVC (Tygon®)	0.76 mm	For sample solution in Inline Dialysis.
6.1826.360	Pump tubing LFL (white/white), 3 stoppers	PVC (Tygon®)	1.02 mm	For sample transfer.
6.1826.380	Pump tubing LFL (gray/gray), 3 stoppers	PVC (Tygon®)	1.25 mm	For Inline Dilution.
6.1826.390	Pump tubing LFL (yellow/yellow), 3 stoppers	PVC (Tygon®)	1.37 mm	For sample solution in Inline Ultrafiltration.

5.12.2.2 Pump tubing connection with filter

The 6.2821.130 filters (42-2) should be changed every 3 months, more frequently at higher backpressure.

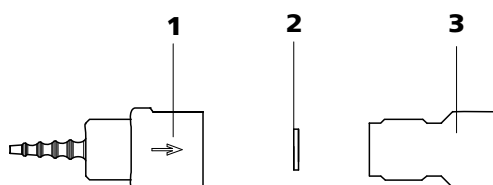


Figure 42 Pump tubing connection – Changing the filter

1 Tubing olive

2 Filter 6.2821.130
Packaging contains 10 items.

3 Filter housing

Replacing the filter

1 Unscrewing filter screw

- Screw the filter screw (42-3) out of the tubing olive (42-1) with the aid of two 6.2621.000 adjustable wrenches.

2 Replacing the filter

- Remove the old filter (42-2) with tweezers.
- Place the new filter (42-2) flat in the tubing olive (42-1) with tweezers.

3 Mounting filter screw

- Screw the filter screw (42-**3**) back into the tubing olive (42-**1**) and tighten by hand. Then additionally tighten with two 6.2621.000 adjustable wrenches.

5.13 Separation column

5.13.1 Separating efficiency

Which analysis quality can be attained, depends to a great extent on the separating efficiency of the separation column used. The separating efficiency of the selected separation column must be sufficient for the analysis problems present. If difficulties occur, you should always first check the quality of the separation column by recording a standard chromatogram.

You can find detailed information on the separation columns available from Metrohm in the leaflet provided along with your separation column, in the **Metrohm IC-Column Program** (available via your Metrohm agent) or in the Internet at <http://www.metrohm.com> in the product area Ion chromatography. You can request free information on special IC applications in the corresponding "**Application Bulletins**" or "**Application Notes**", which are available in the Internet at <http://www.metrohm.com> in the Applications area or via the Metrohm agent responsible.

5.13.2 Protection

To protect the separation column against foreign particles, which can affect the separating efficiency, we recommend that both the eluent and the samples undergo a microfiltration (filter 0.45 µm) before being aspirated via the aspiration filter (6.2821.090).

We recommend always to use a guard column (*see chapter 3.20, page 56*). This protects the separation column and considerably increases its service life. Information regarding which guard column is suitable for your separation column can be found in the **Metrohm IC Column Program** (which is available from your Metrohm agent), the leaflet provided along with your separation column, the product information on the separation column at <http://www.metrohm.com> (product area Ion Chromatography) or obtained directly from your agent.

The pulsation absorber (*see chapter 3.12, page 39*) must be installed in order to protect the column material from pressure concussion caused by injection.

Always store the separation columns sealed and filled according to the data of the column manufacturer when not using them.

The regeneration is considered as the last measure, and not to be carried out regularly.

If the separating properties of the column have deteriorated, the column can be regenerated according to the specifications of the column manufacturer. In the case of separation columns available from Metrohm, the specification for regeneration can be found on the leaflet provided along with each column.

6 Troubleshooting

6.1 Problems and their solutions

Problem	Cause	Remedy
Marked drop in pressure.	<i>Leak in the system.</i>	Check all capillary connections and seal leaks, if necessary (<i>see chapter 3.5, page 16</i>).
The baseline has a large amount of noise.	<i>High pressure pump – contaminated pump valves.</i>	Clean pump valves (<i>see chapter 5.5.2, page 67</i>).
	<i>Eluent – Leakage in eluent path.</i>	Check eluent path.
	<i>Eluent – Blockage in eluent path.</i>	Check eluent path.
	<i>High pressure pump – defective piston seals.</i>	Replace (<i>see chapter 5.5.2, page 67</i>) piston seals .
	<i>Pulsation damper not connected.</i>	Connect the pulsation damper (<i>see Chapter 3.12, Page 39</i>).
	<i>Pulsation absorber not connected. or defective.</i>	Connect pulsation absorber (<i>see chapter 3.12, page 39</i>). or replace it.
The baseline is drifting.	<i>Thermal equilibrium not yet attained.</i>	Condition instrument with the column thermostat (<i>see Chapter 3.15, Page 44</i>) switched on .
	<i>Leak in the system.</i>	Check all capillary connections and seal leaks, if necessary (<i>see chapter 3.5, page 16</i>).
	<i>Eluent – Evaporation of organic solvent in eluent.</i>	■ Check the eluent bottle cap (<i>see figure 13, page 30</i>). ■ Stir the eluent.
The pressure in the system markedly increases.	<i>Inline filter (6.2821.120) blocked.</i>	Replace the filter (6.2821.130) (<i>see chapter 5.6, page 77</i>).
	<i>MSM-HC – blocked.</i>	■ Regenerate the MSM-HC (<i>see chapter 5.11.3.1, page 82</i>). Note: 6.2821.180 pump tubing connection with filter must be used .

Problem	Cause	Remedy
	<i>Guard column – blocked.</i>	Replace guard column (see chapter 3.20, page 56).
	<i>Separation column – blocked.</i>	- Regenerate separation column (see chapter 5.13.4, page 91). - Replace separation column (see "Connecting and rinsing the separation column", page 59). Note: Samples should always be microfiltered (see chapter 5.7, page 79).
	<i>Injection valve – valve blocked.</i>	Have the valve cleaned (by Metrohm service technicians).
The retention times in the chromatogram have changed unexpectedly.	<i>Separation column – diminished separating efficiency.</i>	- Regenerate separation column (see chapter 5.13.4, page 91). - Replace separation column (see "Connecting and rinsing the separation column", page 59).
	<i>Eluent – Gas bubbles in eluent.</i>	- Check the connectors of the eluent degasser (see chapter 3.9, page 32). - Deaerate high-pressure pump (see chapter 3.10.2, page 36).
	<i>High pressure pump – defective.</i>	Request Metrohm Service.
Peak areas are lower than expected.	<i>Sample – leak in the sample path.</i>	Check the sample path.
	<i>Sample – blockage in the sample path.</i>	Check the sample path.
	<i>Sample – sample loop not (completely) filled.</i>	Prolong the sample transfer time.
	<i>Sample – gas bubbles in the sample.</i>	Use sample degasser (see chapter 3.13, page 40).
The peristaltic pump is pumping too little.	<i>Peristaltic pump – contact pressure too weak.</i>	Correctly set contact pressure (see "Set flow rate", page 54).
	<i>Peristaltic pump – filter blocked.</i>	Replace the filter (see chapter 5.12.2.2, page 89).

Problem	Cause	Remedy
Baseline increasing strongly.	<i>MSM-HC – reduced capacity.</i>	Regenerate the MSM-HC (see chapter 5.11.3.1, page 82).
Chromatograms have poor resolution	<i>Separation column – diminished separating efficiency.</i>	- Regenerate separation column (see chapter 5.13.4, page 91). - Replace separation column (see "Connecting and rinsing the separation column", page 59).
Extreme spread of the peaks in the chromatogram. Splitting (dual peaks)	<i>Capillary connections – dead volume in the system.</i>	Check connections (see chapter 3.5, page 16) (use PEEK capillaries with an internal diameter of 0.25 mm between the injection valve and detector).
	<i>Guard column – diminished performance.</i>	Replace guard column (see chapter 3.20, page 56).
	<i>Separation column – dead volume at column head.</i>	- Install the separation column in reverse flow direction (if permitted by the leaflet) and rinse into a beaker. - Replace separation column (see "Connecting and rinsing the separation column", page 59).
Precision problems - the measured values are highly scattered.	<i>Sample – gas bubbles in the sample.</i>	Use sample degasser (see chapter 3.13, page 40).
	<i>Injection valve – sample loop.</i>	Check installation of the sample loop (see chapter 3.14.1, page 42).
	<i>Sample – rinsing volume too low.</i>	Increase rinsing time (see chapter 5.8, page 79).
	<i>Injection valve – defective.</i>	Request Metrohm Service.
Vacuum is not being built	<i>Eluent degasser – Vacuum connector on the rear of the instrument not (tightly) sealed.</i>	Seal the Vacuum connector tightly with a threaded stopper (6.1446.040).

7.5 Housing

Dimensions

Width	365 mm
Height	642 mm
Depth	380 mm

Material of base tray, housing and flask holder	Polyurethane hard foam (PUR) with flame retardation for fire class V0, CFC-free, painted
---	--

Operating elements

Indicators	LED for power display
On/Off switch	On the rear of the instrument

7.6 Eluent degasser

Material	Fluoropolymer
Resistance to solvents	No restriction (except PFC)
Time to establish vacuum	< 60 s

7.7 High pressure pump

Type	▪ Serial dual-piston pump ▪ Intelligent pump head recognition ▪ Chemically inert ▪ Metal-free pump heads ▪ Materials in contact with the eluent: PEEK, ZrO₂, PTFE/PE ▪ Self-optimizing flow and pressure
------	--

Flow rate

Adjustable flow range	0.001...20.0 mL/min
Flow increment	1 µL/min
Reproducibility of the eluent flow	< 0.1 % deviation

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